

Control of estrus and ovulation in beef heifers¹

D.J. Patterson, J.M. Thomas, B.E. Bishop, J.M. Abel, J.W.C. Locke, and M.F. Smith, Division of Animal Sciences, University of Missouri, Columbia, MO

Introduction

Improving traits of major economic importance in beef cattle can be accomplished most rapidly through selection of genetically superior sires and widespread use of artificial insemination. Estrus synchronization and artificial insemination (AI) remain the most important and widely applicable reproductive biotechnologies available for cattle (Seidel, 1995); and in recent years, the development of convenient and economical protocols to synchronize estrus and ovulation to facilitate use of fixed-time AI (FTAI) with resulting high fertility resulted in increased adoption of these important management practices.

Procedures that facilitate synchronization of estrus in estrous cycling females and induction of an ovulatory estrus in peripubertal heifers and anestrus postpartum cows will increase reproductive rates and expedite genetic progress. Estrus synchronization can be an effective means of increasing the proportion of females that become pregnant early in the breeding season resulting in shorter calving seasons and more uniform calf crops (Dziuk and Bellows, 1983). Females that conceived to a synchronized estrus calved earlier in the calving season and weaned calves that were on average 13 days older and 21 pounds heavier than calves from non-synchronized females (Schafer et al., 1990). Effective estrus synchronization programs offer the following advantages: 1) cows or heifers are in estrus at a predicted time which facilitates AI, embryo transfer, or other assisted reproductive techniques; 2) the time required for detection of estrus is reduced thus decreasing labor expense associated with estrus detection; 3) AI becomes more practical, enabling producers to utilize genetically superior high accuracy sires; 4) cattle will conceive earlier during the breeding period; and 5) calves will be older and heavier at weaning.

Until recently, the inability to predict time of estrus for individual cows or heifers in a group often made it impractical to use AI because of the labor required for detection of estrus. However, recent improvements in methods to control estrus and ovulation in beef cows and heifers, now provides the opportunity to expand the use of AI without the need to detect estrus. Procedures to control the estrous cycle of the cow can improve reproductive rates and speed up genetic progress. These procedures include synchronization of estrus in estrous cycling females, and induction of estrus accompanied by ovulation in heifers that have not yet reached puberty or among cows that have not returned to estrus after calving.

The following protocols and terms will be referred to throughout this manuscript.

Protocols for AI performed on the basis of detected estrus:

PG: Prostaglandin F_{2α} (PG; Lutalyse[®], Estrumate[®], ProstaMate[®], InSynch[®], estroPLAN[®]).

MGA-PG: Melengestrol acetate (MGA; 0.5 mg/hd/day) is fed for a period of 14 days with PG administered 17 to 19 days after MGA withdrawal.

GnRH-PG (Select Synch): Gonadotropin-releasing hormone injection (GnRH; Cystorelin[®], Factrel[®], Fertagyl[®], OvaCyst[®]) followed in 7 days with an injection of PG.

¹Acknowledgements. Research summarized in this manuscript was supported by National Research Initiative Competitive Grant no. 00-35203-9175 and 2005-55203-15750 from the USDA National Institute of Food and Agriculture. The authors gratefully acknowledge Zoetis (Florham Park, NJ) for providing Lutalyse and EAZI BREED CIDR Cattle inserts; Merial (Athens, GA) for providing Cystorelin; Bayer (Monheim, Germany) for providing OvaCyst; ABS Global, Accelerated Genetics, Genex Cooperative, Inc., and Select Sires, Inc., for providing semen. We express appreciation as well to Circle A Ranch, Lineville, IA; SL Lock at Genex Cooperative, Inc.; Mason-Knox Ranch, Frankfort, SD; Ogren Angus, Langford, SD; and DS McAtee and JJD Schreffler at the University of Missouri Thompson Research Center for their dedicated support of this research.

MGA-GnRH-PG (MGA® Select): MGA is fed for 14 days, GnRH is administered 12 days after MGA withdrawal, and PG is administered 7 days after GnRH.

CIDR Select: CIDRs are inserted on day 0 and removed on day 14, GnRH is administered on day 23 and PG is administered on day 30.

14-day CIDR-PG: CIDRs are inserted on day 0 and removed on day 14. PG is administered on day 30.

Protocols for fixed-time AI in beef heifers:

MGA-PG: Melengestrol acetate (MGA; 0.5 mg/hd/day) is fed for a period of 14 days with PG administered 19 days after MGA withdrawal. Insemination is performed 72 hours after PG, with GnRH administered at AI.

7-day CO-Synch + CIDR: GnRH is administered at CIDR insertion on day 0, followed 7 days later with CIDR removal, and PG. Insemination is performed 54 hours after CIDR removal and PG, with GnRH administered at AI.

5-day CO-Synch + CIDR: GnRH is administered at CIDR insertion on day 0, followed 5 days later with CIDR removal, and PG. A second injection of PG is administered 8 hours after CIDR removal and the first PG injection. Insemination is performed 60 hours after CIDR removal and the first injection of PG, with GnRH administered at AI.

CIDR Select: CIDRs are inserted on day 0 and removed on day 14, GnRH is administered on day 23 and PG is administered on day 30. Insemination is performed 72 hours after PG with GnRH administered at AI.

14-day CIDR-PG: CIDRs are inserted on day 0 and removed on day 14 with PG administered on day 30. Insemination is performed 66 hours after PG with GnRH administered at AI.

Terms:

Estrous response: The number of females that exhibit estrus during a synchronized period.

Synchronized period: The period of time during which estrus is expressed after treatment.

Synchronized conception rate: The proportion of females that became pregnant of those exhibiting estrus and inseminated during the synchronized period.

Synchronized pregnancy rate: Proportion of females that become pregnant of the total number treated.

To avoid problems when using estrus synchronization, heifers should be selected for a program when the following conditions are met: 1) replacement heifers are developed to prebreeding target weights based on a specified percentage of the heifer's projected mature weight; and 2) reproductive tract scores (RTS) are assigned to heifers no more than two weeks before a synchronization treatment begins [scores of 2 or higher on a scale of 1 to 5] and at least 50 percent of the heifers are assigned a RTS of 4 or 5 (Patterson et al., 2000a).

Estrus synchronization and artificial insemination contribute to a total heifer development program

Estrus synchronization and artificial insemination contribute to a total heifer development program in several ways. Estrus synchronization improves time management for producers that use AI by

concentrating the breeding and resulting calving periods. Managers are able to spend more time observing heifers as they calve because calving occurs over a shorter time period. Calf losses in many cases are reduced because of improved management during the calving period. Artificial insemination provides the opportunity to breed heifers to bulls selected for low BW or high calving ease EPD with high accuracy. This practice minimizes the incidence and severity of calving difficulty and decreases calf loss that results from dystocia. In addition, heifers that conceive during a synchronized period typically wean calves that are older and heavier at weaning time (Schafer et al., 1990). Finally, heifer calves that result from AI can be an excellent source of future replacements facilitating more rapid improvement in the genetic makeup of an entire herd.

Progestins were used to induce estrus in peripubertal heifers (Gonzalez-Padilla et al., 1975) and were originally combined with estrogen to mimic changes that occur in concentrations of blood hormones around the time of puberty. Increased progesterone is thought to be a prerequisite for the development of normal estrous cycles. Progesterone increases during the initiation of puberty in the heifer (Berardinelli et al., 1979), and before resumption of normal ovarian cyclicity in postpartum suckled beef cows (Prybil and Butler, 1978; Rawlings et al., 1980). Progestins stimulate an increase in follicular growth that results subsequently in increased production of estrogen by ovarian follicles (Henricks et al., 1973; Wetteman and Hafs, 1973; Sheffel et al., 1982; Garcia-Winder et al., 1986). Melengestrol acetate and CIDR initiate estrous cyclicity in peripubertal beef heifers (Patterson et al., 1990; Lucy et al., 2001) and are associated with increased LH pulse frequency during the treatment period (Smith and Day, 1990; Imwalle et al., 1998). Recent studies suggest that the stimulatory effects of progestins on LH secretion are greatest after removal of the steroid (Hall et al., 1997; Imwalle et al., 1998). Furthermore, improvements in observed pubertal induction response following treatment with a progestin occur with an increase in age (Hall et al., 1997). The increase in pulsatile release of LH that occurs in response to progestin treatment in peripubertal heifers results in a decrease in estrogen receptors within neuronal systems that mediate negative feedback actions of estradiol on GnRH secretion (Anderson et al., 1996).

Burfening (1979) suggested that because puberty is a heritable trait, induced puberty in replacement heifers over several generations might result in situations in which attainment of puberty would be difficult without hormone treatment. This consideration cannot be overlooked. However, there is a need to explore treatments to induce puberty in breeds of cattle that are late-maturing but of sufficient age and weight at the time of treatment to permit successful application (Patterson et al., 1990). The decision to utilize this practice within a herd perhaps differs with various types of beef operations. For instance, the common goal of most managers of commercial cow-calf herds is to maximize weaning rate. In other words, the investment in time and resources in a heifer from weaning to breeding requires that management efforts be made to facilitate puberty onset and maximize the likelihood of early pregnancy. In this scenario, a method to induce puberty in heifers could serve as a valuable tool to improve reproductive performance of heifers retained for breeding purposes. On the other hand, seed stock managers should weigh the economic importance of puberty onset in their herds, as well as their customers', and the associated potential and resulting implication of masking its true genetic expression.

MGA-based programs

This review includes methods to control estrous cycles of cattle using MGA. Four methods are outlined for using the MGA program to facilitate estrus synchronization in beef heifers. The choice of which system to use depends largely on a producer's goals. Melengestrol acetate is the common denominator in each of the four systems presented here. Melengestrol acetate is an orally active progestin. When consumed on a daily basis, MGA will suppress estrus and prevent ovulation (Imwalle et al., 2002). Melengestrol acetate may be fed with a grain or a protein carrier and either top-dressed onto other feed or batch mixed with larger quantities of feed. Melengestrol acetate is fed at a rate of 0.5 mg/animal/day in a single daily feeding.

The duration of feeding may vary among protocols, but the level of feeding is consistent and critical to success. Animals that fail to consume the required amount of MGA on a daily basis may prematurely return to estrus during the feeding period. This can be expected to reduce the estrous response during the synchronized period. Therefore, adequate bunk space (60 linear cm/head) must be available so that all animals consume feed simultaneously (Patterson et al., 2003). Animals should be observed for behavioral signs of estrus each day of the feeding period. This may be done as animals approach the feeding area and before feed distribution. This practice will ensure that all females receive adequate intake. Heifers will exhibit estrus beginning 48 hours after MGA withdrawal, and this will continue for 6 to 7 days. It is generally recommended that females exhibiting estrus during this period not be inseminated or exposed for natural service because of reduced fertility females experience at the first heat after MGA withdrawal.

Method 1: MGA with natural service.

The simplest method involves using bulls to breed synchronized groups of females. This practice is useful in helping producers make a transition from natural service to artificial insemination. In this process, heifers receive the normal 14-day feeding period of MGA and are then exposed to fertile bulls about 10 days after MGA withdrawal (Figure 1).

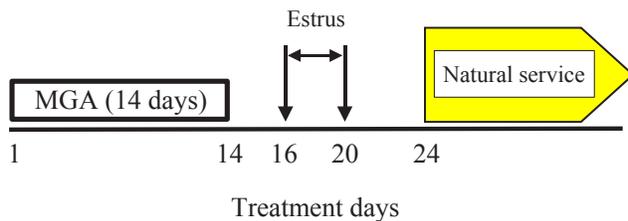


Figure 1. MGA and natural service (adapted from Patterson et al., 2000b).

This system works effectively, however careful consideration of bull to female ratios is advised. It is recommended that 15 to 20 synchronized females be exposed per bull. Age and breeding condition of the bull and results of breeding soundness examinations should be considered.

Method 2: MGA + Prostaglandin.

This method of estrus synchronization involves the combination of MGA with prostaglandin $F_{2\alpha}$. Prostaglandin $F_{2\alpha}$ (PG) is a luteolytic compound normally secreted by the uterus of the cow. Prostaglandin $F_{2\alpha}$ can induce luteal regression but cannot inhibit ovulation. When PG is administered in the presence of a functional corpus luteum (CL) during days 6 to 16 of the estrous cycle, premature regression of the CL begins and the cow returns to estrus. In this program, prostaglandin should be administered 19 days after the last day of MGA feeding. This treatment places all animals in the late luteal stage of the estrous cycle at the time of PG injection, which shortens the synchronized period and maximizes conception rate (Figure 2). Although a 19-day interval is optimal, 17- to 19-day intervals produce acceptable results and provide flexibility for extenuating circumstances (Brown et al., 1988; Deutscher, 2000; Lamb et al., 2000). Five available PG products for synchronization of estrus in cattle can be used after the MGA treatment: Estrumate[®], estroPLAN[®], InSynch[®], Lutalyse[®], or ProstaMate[®]. Label-approved dosages differ with each of these products; carefully read and follow directions for proper administration before their use.

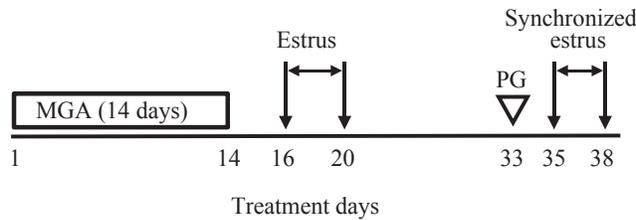


Figure 2. The MGA-PG protocol (adapted from Brown et al., 1988; Deutscher, 2000; Lamb et al., 2000).

Management considerations related to long-term feeding of MGA to heifers. Long-term feeding of MGA to beef heifers and associated effects on fertility may be a concern in specific production systems. It is not uncommon for heifers to be placed on MGA for extended periods of time and subsequently exposed for breeding after placement in backgrounding programs that necessitate long-term MGA administration. Zimbelman et al. (1970) reported no negative effect of either long-term or repeated intervals of feeding MGA to beef heifers and cows, other than the expected reduced conception rate when cattle were bred at the synchronized estrus 3 to 7 days after the last day of MGA feeding. Patterson et al. (1993) designed a study (Figure 3) to compare estrous response and fertility during synchronized estrous periods among beef heifers that were fed MGA for 87 days (long-term, LT) or 14 days (short-term, ST) prior to PG. Heifers were stratified by age and weight to LT- or ST-MGA treatments (Table 1), and received 0.5 mg MGA per head per day for 87 or 14 days, respectively. Heifers in each group were administered PG 17 days after MGA withdrawal. Heifers in both groups that failed to exhibit estrus within 6 days after the first injection of PG, were administered a second injection of PG 11 days later (Figure 3).

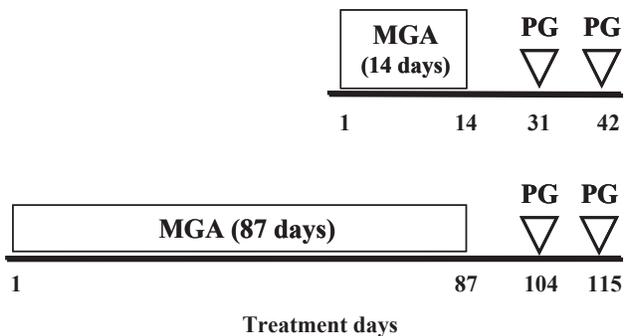


Figure 3. Comparison of short-term and long-term MGA treatments.

Transrectal ultrasonography was used to examine ovaries of all heifers at the end of treatment with MGA and at the time PG was administered. Heifers that failed to exhibit estrus after the first injection of PG were re-examined prior to the second PG injection. All heifers were exposed for natural-service for an additional 45 d after the AI period. More ST-treated heifers exhibited estrus after the first injection of PG than LT-treated heifers (Table 2; $P < 0.05$). Total response after the two injections of PG, however, did not differ between treatments. Furthermore, there were no significant differences between treatments in synchronized conception or pregnancy rates, or pregnancy rates at the end of the breeding period (Table 2). A higher incidence of luteinized follicular cysts (Table 3) was observed among heifers in the LT-treatment compared with heifers in the ST-treatment [LT, 11/30 (37%); ST, 0/31 (0%)]. This observation may explain differences in estrous response between treatments following the first injection of PG. These data indicate that long-term feeding of MGA may result in a higher than normal incidence of luteinized follicular cysts and an

associated reduction in estrous response after PG. The data indicate, however, that re-injection with PG resulted in satisfactory breeding performance among heifers that were fed MGA for extended periods of time.

Table 1. Ages and weights of heifers at the time PG was administered.

Treatment	No. of heifers	Age, d	Weight, lb
Short-term, 14 d	31	427	865
Long-term, 87 d	30	423	851

¹Adapted from Patterson et al., 2003.

Table 2. Estrous response and fertility of heifers treated long-term or short-term with MGA.

Response variable	Short-term MGA, 14 d			Long-term MGA, 87 d		
	1st PG ^a	2nd PG ^a	Total	1st PG ^a	2nd PG ^a	Total
Estrous response	24/31 (77% ^b)	4/7 (57%)	28/31 (90%)	16/30 (53% ^c)	10/14 (71%)	26/30 (87%)
Synchronized conception	15/24 (63%)	3/4 (75%)	18/28 (64%)	12/16 (75%)	6/10 (60%)	18/26 (69%)
Synchronized pregnancy	-----	18/31 (58%)	-----	18/30 (60%)		
Final pregnancy	-----	28/31 (90%)	-----	27/30 (90%)		

^a 1st PG refers to animals that responded to PG administered 17 days after MGA withdrawal. 2nd PG refers to animals that failed to respond to the first injection of PG that were reinjected 11 days later.

^{b, c} Percentages within row and between treatments with unlike superscripts differ ($P < 0.05$; Adapted from Patterson et al., 2003).

Table 3. Ovarian morphology of heifers treated long-term or short-term with MGA.

Treatment	Normal	Abnormal ^a
Short-term	31/31 (100%)	0/31 (0%)
Long-term	19/30 (63%)	11/30 (37%)

^a Abnormal = presence of luteinized follicular cysts, 20-45 mm diameter (Adapted from Patterson et al., 2003).

Method 3: MGA[®] Select

Studies with heifers showed that both synchrony of estrus and total estrous response were improved when PG is administered 19 days after MGA withdrawal compared with those of heifers injected on day 17 after MGA withdrawal (Deutscher et al., 2000; Lamb et al., 2000). Based on these data, we evaluated a modified MGA-PG protocol for inducing and synchronizing a fertile estrus in beef heifers (Wood et al., 2001; Figure 4). The first modification changed the day of PG injection from day 31 to day 33 of treatment. The second modification was the addition of GnRH on day 26 of treatment. We found that the addition of GnRH on day 26 of the MGA-PG protocol induced luteal tissue formation and initiated a new follicular wave on approximately day 28 in cycling beef heifers (Figure 5B). The proportion of heifers with synchronized follicular waves on day 33 was increased significantly compared to heifers that did not receive GnRH (Wood et al., 2001; Figure 5A and 5B).

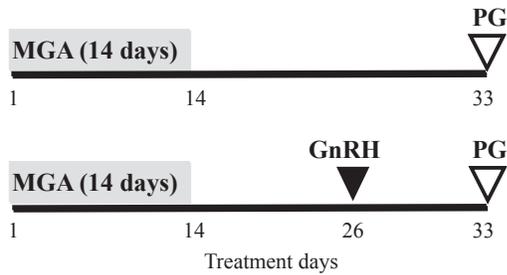


Figure 4. A modified long-term MGA protocol. Heifers were fed MGA for 14 days; 19 days after MGA withdrawal PG was administered to all heifers. GnRH was administered to ½ of the heifers 7 days prior to PG (Wood et al., 2001).

Wood-Follis et al. (2004) reported differences in estrous response and synchrony of estrus during the synchronized period among heifers assigned to the treatments illustrated in Figure 6A,B. This difference in estrous response and degree of synchrony was based on the percentage of heifers that were pubertal at the time treatment with MGA began. Figures 6A and 6B illustrate these differences (Wood-Follis et al., 2004). Figure 6A shows the distribution of estrus where only 30% of the heifers were pubertal at the time treatment with MGA began, whereas Figure 6B illustrates the distribution of estrus for heifers where 56% of the heifers were pubertal at the same time. The increased degree of estrous cyclicality of heifers shown in Figure 6B was associated with a reduced variance in the interval to estrus among MGA-GnRH-PG treated heifers. AI pregnancy rates remained high for both MGA-GnRH-PG and MGA-PG treated heifers and were not different (67% and 60%, respectively [Figure 6A] and 75% and 72%, respectively [Figure 6B]).

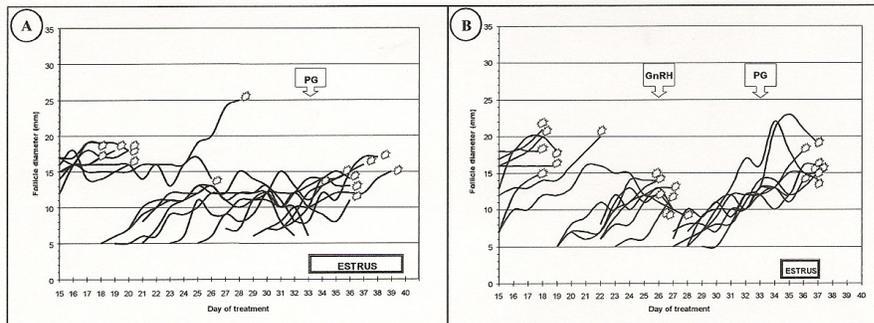


Figure 5A and 5B. Patterns of dominant follicle development in control (MGA-PG; A) and GnRH treated (MGA-GnRH-PG; B) heifers. Administration of GnRH (B) caused the synchronized development of a dominant follicle before PG injection. Follicular development in MGA-PG treated heifers was poorly synchronized (Wood et al., 2001).

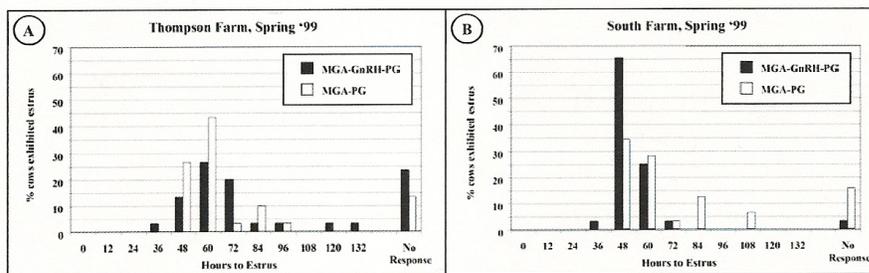


Figure 6A and 6B. Percentage of heifers observed in estrus for MGA-PG and MGA-GnRH-PG treated heifers. Estrous cyclicality rates were 30% and 56% for heifers at Location 1 (A) and 2 (B), respectively, at the time treatment with MGA began (Wood-Follis et al., 2004).

Method 4: MGA[®] - PG with fixed-time AI. This method involves the combination of MGA with prostaglandin F_{2α} as previously described, however in this case fixed-time AI is performed 72 hours after PG with GnRH administered at AI (Figure 7).

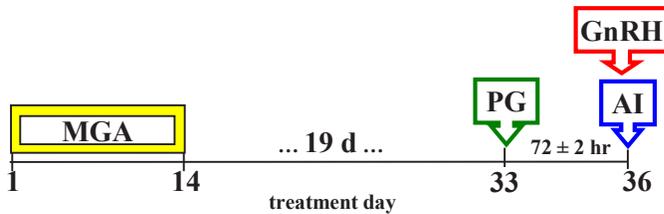


Figure 7. MGA[®]- PG with fixed-time AI 72 hours after PG.

Additional considerations. An additional consideration for Methods 2 and 3 (MGA-PG and MGA Select) pertains to heifers that fail to exhibit estrus after the last PG injection. In this case, non-responders may be re-injected with PG 11 to 14 days after the last injection of PG was administered. These females would then be observed for signs of behavioral estrus for an additional 6 to 7 days. This procedure would maximize efforts to inseminate as many females within the first 2 weeks of the breeding period as possible. Females that were inseminated during the first synchronized period should not be re-injected with PG. In addition, the decision to use Method 3 in heifers should be based on careful consideration of the heifer's age, weight, and pubertal status (Zimelman, 1963; Zimelman and Smith, 1966; Patterson et al., 1989; Federal Register, 1997; Wood-Follis et al., 2004).

Development of the 7-day CIDR-PG protocol for heifers

Lucy et al. (2001; Table 4) summarized results from initial studies conducted in the U.S. involving controlled internal drug release (CIDR)-based protocols for use in synchronizing estrus in beef heifers. These data were submitted to FDA in support of the original approval for the CIDR in beef heifers and cows. Three treatments were involved in the study and included: 1) an untreated control; 2) PG only; and 3) 7-day CIDR-PG. The 7-day CIDR-PG treated heifers had CIDRs inserted for 7 days with PG administered on day 6 of CIDR treatment. The 7-day CIDR-PG protocol yielded greater pregnancy rates compared with control or PG treated heifers. Treatment with CIDR increased synchronization rates within the first 3 d following PG, resulting in enhanced pregnancy rates. The improved pregnancy rate in prepubertal beef heifers treated with the CIDR was noteworthy because prepubertal heifers in the control or PG treatments never attained pregnancy rates that were similar to those of the 7-day CIDR-PG treated heifers. The drawback of the protocol was that PG was administered on d 6 after CIDR insertion, which required an additional day of handling the heifers.

Table 4. Synchronization, conception, and pregnancy rate for beef heifers (modified from Lucy et al, 2001).

Item	Synchronization rate		Conception rate		Pregnancy rate	
	No.	%	No.	%	No.	%
Prepubertal						
Control	8/107	7	6/8	75	6/7	6
PG	11/101	11	6/11	50	6/101	6
CIDR-PG	50/105	48	29/50	58	29/105	28
Cyclic						
Control	25/44	17	13/25	52	13/144	9
PG	56/151	37	29/56	52	29/151	19
CIDR-PG	93/116	80	57/93	61	57/116	49
Total						
Control	33/151	22	19/33	58	19/151	13
PG	67/252	27	35/67	52	35/252	14
CIDR-PG	143/221	65	86/143	60	86/221	39

Dejarnette (unpublished data) compared timing of estrus, estrous response and pregnancy rate resulting from AI during the synchronized period among heifers that were assigned to a 7-day CIDR treatment and that received PG on day 6 or 7 of the treatment schedule. Although heifers that received PG on day 6 (one day prior to CIDR removal) exhibited estrus earlier than heifers that received PG on day 7 (at CIDR removal), there were no differences between groups for the response variables considered. Therefore, to simplify treatment administration, PG is in most cases now administered coincident with the time of CIDR removal.

The multi-state CIDR trial. Lamb et al. (2006) lead a multi-state effort involving 12 locations in 6 states to determine whether: 1) administration of an estrus synchronization protocol followed by FTAI could yield pregnancy rates similar to a protocol requiring detection of estrus; and 2) whether an injection of GnRH at CIDR insertion enhanced pregnancy rates in beef heifers. Four treatments were involved in the study (Figure 8). Heifers in treatments 1 and 2 were handled in the same way, with the exception of GnRH administration at CIDR insertion. Heifers in both treatments were observed for signs of behavioral estrus and inseminated on the basis of observed estrus up through 72 h after PG. Eighty four hours after the administration of PG all heifers that failed to exhibit estrus to that point were inseminated by appointment with GnRH administered at AI. Heifers in treatments 3 and 4 received the same treatment schedules as heifers in treatments 1 and 2, respectively however heifers in both treatments 3 and 4 were inseminated by appointment 60 hours after PG with GnRH administered at AI. Although no differences in pregnancy rates were detected among treatments, heifers that were inseminated in the estrus-detection treatments had numerically higher pregnancy rates than heifers in the fixed-time AI treatments (Table 5).

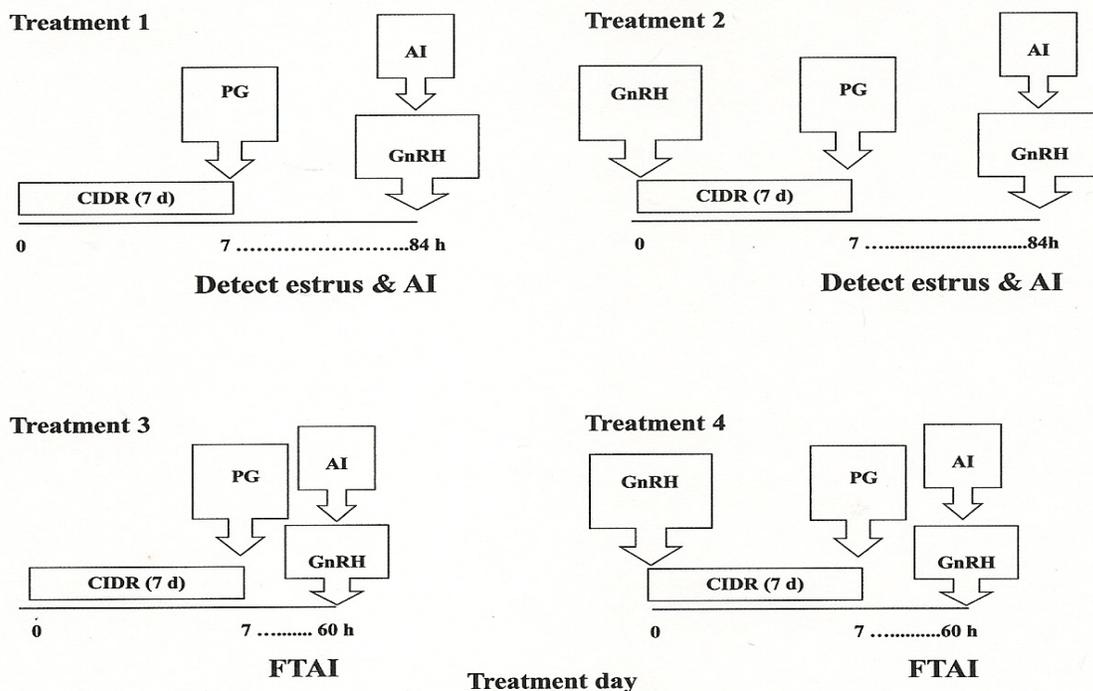


Figure 8. Treatment schedules for heifers in the multi-state CIDR trial (Lamb et al., 2006).

Table 5. Pregnancy rates following AI among beef heifers in the multi-state CIDR trial (Lamb et al, 2006).¹

Item	Treatments							
	1		2		3		4	
	No.	%	No.	%	No.	%	No.	%
Prepubertal	19/36	53	32/54	59	22/36	61	28/44	64
Cycling	195/341	57	201/317	63	189/353	54	185/346	54

¹Refer to Figure 7 for a description of the 4 treatment protocols.

How do MGA- and CIDR-based protocols compare?

Substituting EAZI-BREED CIDR inserts for MGA in the MGA Select protocol in beef heifers.

Utter and Corah (1994) reported cases in which pregnancy rates resulting from MGA-based estrus synchronization protocols are lower than expected in yearling age heifers. These instances of reduced fertility were generally associated with heifers in which estrous cyclicity rates were high, and heifers generally weighed more and were in higher body condition prior to treatment with MGA compared to lighter weight, lower body conditioned heifers. Based on these considerations, Kojima et al. (2004) designed a study to compare pre-synchronization with MGA or CIDR (14-day treatment), followed 12 or 9 d later, respectively, with an injection of GnRH, and PG 7 d after GnRH. The treatments were compared on the basis of estrous response, timing of AI, and pregnancy rate in beef heifers. No differences in estrous response were detected between MGA Select and 14-d CIDR treated heifers; however, 14-d CIDR treated heifers showed an improvement in synchrony of estrus, conception, and pregnancy rates during the synchronized period. These improvements associated with a 14-d CIDR treatment were attributed to a reduced interval to estrus (Macmillian and Peterson, 1993) and improved synchronization of follicular waves after CIDR removal as compared to the end of MGA feeding.

A widely held hypothesis is that GnRH is less effective at synchronizing follicular waves in heifers compared to cows. Lamb et al. (2006) reported no difference in synchrony of estrus or pregnancy rate between CIDR + PG and Select Synch + CIDR treated heifers, suggesting that response to GnRH in heifers at CIDR insertion may be of limited value. Atkins et al. (2008; Table 6) evaluated follicular response to GnRH among pubertal beef heifers on specific days of the estrous cycle. Response was based on ovulation or luteinization of a dominant follicle and subsequent initiation of a new follicular wave in response to GnRH. These data (Table 6) support the concept that presynchronization prior to initiation of the GnRH + PG protocol may be of greater importance in heifers, and therefore significant in relation to success we initially reported with the long-term CIDR-GnRH-PG protocol (Kojima et al., 2004).

Schafer et al. (2006) characterized follicular dynamics, timing of estrus, and response to GnRH in yearling beef heifers after treatment with the 14-day CIDR protocol. The objective of the experiment was to characterize response after treatment with a 14-day CIDR insert followed by the administration of GnRH and PG in 79 Angus crossbred heifers. At the initiation of the experiment 53 heifers were estrous cycling and 26 were prepubertal based on two blood samples for progesterone collected 10 days and 1 day prior to initiation of treatment. CIDRs were inserted into all heifers on the same day for 14 days, GnRH was injected on day 23, and PG on day 30. Estrus detection was performed continuously after CIDR removal using the HeatWatch® Estrus Detection System. The study characterized estrous response and timing of estrus after treatment with the 14-day CIDR, follicular dynamics the day preceding and the day GnRH was administered, response to GnRH, and timing of estrus after PG. Sixty-nine heifers exhibited estrus (47 pubertal, 22 prepubertal) after CIDR removal.

Table 6. Response to GnRH in estrous cycling beef heifers based on the day of the estrous cycle GnRH was administered (From Atkins et al., 2008).

Day of treatment	1st GnRH (no. & % responding)	2nd GnRH (no. & % responding)
Day 2	0/7 = 0%	3/7 = 43%
Day 5	8/8 = 100%	8/8 = 100%
Day 10	0/6 = 0%	5/6 = 83%
Day 15	5/8 = 63%	1/8 = 13%
Day 18	5/8 = 63%	2/8 = 25%

There was no difference ($P > 0.05$) in the interval to estrus after CIDR removal for pubertal and prepubertal heifers [50.0 ± 27.3 pubertal, and 48.1 ± 28.3 h prepubertal, respectively]. Follicular dynamics were recorded for all heifers the day preceding GnRH, the day GnRH was administered, and resulting response to GnRH. Comparisons were made on the basis of the day of the estrous cycle heifers were on at the time GnRH was administered based on the day estrus was expressed after CIDR removal. There was a significant effect ($P < 0.05$) of day of the estrous cycle on mean follicle diameter at the time GnRH was administered. Response to GnRH was highest among heifers with dominant follicles ≥ 10.0 mm (64/71, 90%) and lower among heifers with follicles < 10 mm (4/8, 44%). Mean follicle diameter was ≥ 10.0 mm among all heifers that were on d 5, 6, 7 or 8 of the estrous cycle at the time GnRH was administered. Concentrations of progesterone in serum at PG were higher among pubertal versus prepubertal heifers [7.9 pubertal versus 6.9 ng/ml prepubertal, respectively]. Estrous response after PG did not differ among pubertal and prepubertal heifers and peaked between 48 and 60 hours. The study provided a descriptive comparison of response to presynchronization with a CIDR prior to GnRH and PG in pubertal and prepubertal beef heifers.

We used this protocol successfully in conjunction with either heat detection and AI (Leitman et al., 2008) or FTAI with AI performed 72 hours after PG and GnRH administered at the time of AI (Busch et al., 2007;

Figure 9). On-farm field trials are summarized in Table 7 reporting results after use of the CIDR Select protocol in conjunction with breeding programs requiring heat detection or fixed-time AI. It is interesting to note that pregnancy rates following administration of the CIDR Select protocol were comparable whether heifers were inseminated on the basis of observed estrus (Table 7) or at predetermined fixed times (Table 7).

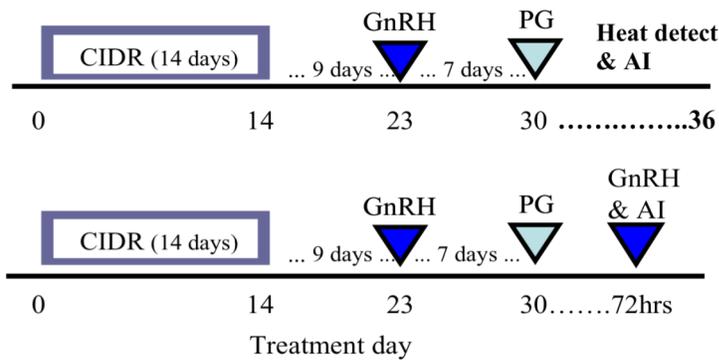


Figure 9. Estrus synchronization schedules involving use of the CIDR Select protocol in breeding programs for beef heifers that require heat detection or fixed-time AI.

Table 7. Pregnancy rates after administration of the CIDR Select protocol in field trials involving AI performed after observed estrus or fixed-time AI performed 72 hours after PG (Patterson et al., 2006).

Breeding program	No. pregnant	No. inseminated	Pregnancy rate (%)
Estrus detection & AI	499	830	60
Fixed-time AI	518	853	61

Tauk et al. (2007) compared CIDR-PG and MGA-PG protocols in beef heifers. The study was designed to compare: 1) estrous synchronization response following progestin removal, and PG administered 17 or 19 days after progestin withdrawal, and b) AI pregnancy rates during the synchronized period. More ($P < 0.05$) CIDR-treated heifers exhibited estrus within 120 h after progestin removal than MGA-treated heifers. Intervals to estrus after progestin removal were shorter ($P < 0.05$) for CIDR-treated heifers than MGA-treated heifers and more ($P < 0.05$) CIDR-treated heifers exhibited estrus and were inseminated within 60 h after PG than MGA-treated heifers. Pregnancy rates did not differ between MGA-treated (66%) and CIDR-treated heifers (62%). Tauk et al. (2007) concluded that use of CIDR as a progestin source was equally effective as MGA in synchronizing estrus in beef heifers.

Mallory et al. (2010) conducted two experiments to evaluate 14-day MGA and CIDR-based estrus synchronization protocols on the basis of potential for use in facilitating FTAI in estrous cycling and prepubertal beef heifers. Heifers in the first experiment (Figure 10) were fitted with HeatWatch® estrus detection transmitters at the time of progestin removal for continuous estrus detection, and in both experiments the synchronized period was designated as 0 to 144 h following PG. HeatWatch transmitters were maintained on all heifers until AI was performed. Figure 11 illustrates the pattern of estrus distribution following withdrawal of MGA from feed or removal of CIDR for the respective treatments. The variance associated with interval to estrus after progestin withdrawal/removal was significantly reduced ($P < 0.01$) among 14-day CIDR-PG compared to MGA-PG treated heifers.

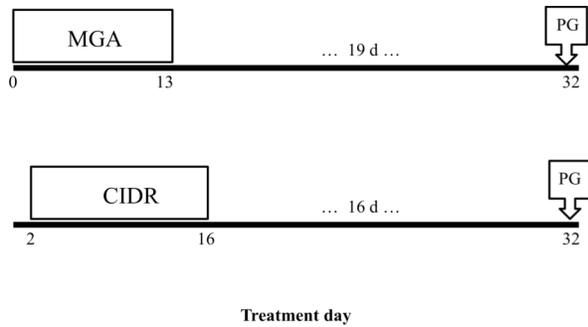


Figure 10. Treatment schedule for heifers assigned to the MGA-PG and 14-day CIDR-PG treatment protocols. (Mallory et al., 2010).

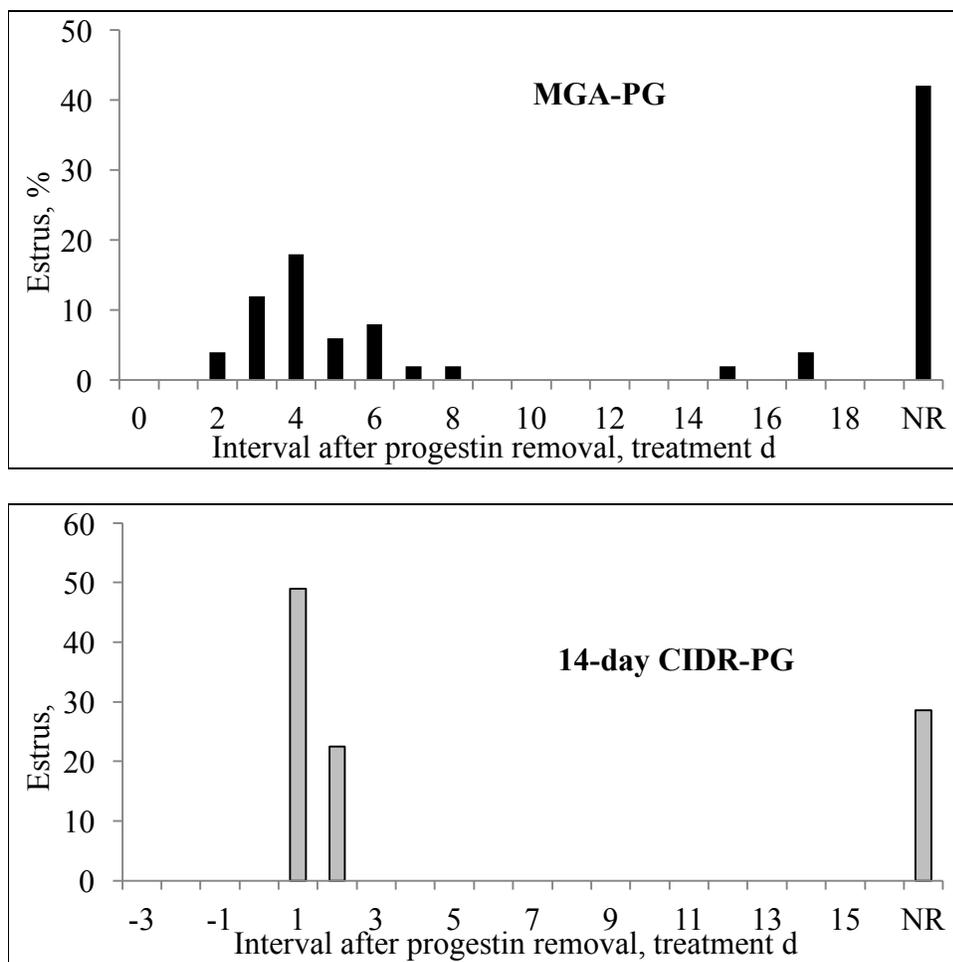


Figure 11. Percentage of heifers in MGA-PG and 14-day CIDR-PG treatments that exhibited estrus after withdrawal/removal of progestin: MGA-PG (black bar) and 14-day CIDR-PG (gray bar). NR = no estrous response. Heifers assigned to MGA-PG received MGA in a 1.0-kg feed supplement for 14 d and were administered PG on d 32. Heifers assigned to 14-day CIDR-PG received a CIDR insert from d 2 of treatment to d 16, and PG on d 32 (Mallory et al., 2010).

Estrous response after PG was greater ($P = 0.01$) for 14-day CIDR-PG (92%) than for MGA-PG (85%) treated heifers (Table 8). The distribution of estrus after PG is depicted in Figure 12. The mean interval to estrus after PG did not differ ($P = 0.73$) between MGA-PG (57.4 ± 2.5 h) and 14-day CIDR-PG (56.2 ± 2.5 h) treated heifers (Table 9). There was however, a significant difference in the mean interval to estrus after PG ($P = 0.04$) between estrous cycling (62.4 ± 2.4 h) and prepubertal heifers (52.4 ± 4.4 h) assigned to the MGA-PG protocol, but no difference ($P = 0.75$) between estrous cycling and prepubertal heifers assigned to the 14-day CIDR-PG protocol (55.4 ± 2.4 h and 57.0 ± 4.4 h, respectively). The variance associated with interval to estrus after PG was reduced ($P < 0.01$) among 14-day CIDR-PG heifers than for MGA-PG treated heifers. Variance for interval to estrus after PG differed between treatments for estrous cycling ($P < 0.01$) and prepubertal ($P < 0.05$) heifers; however, variance for interval to estrus after PG did not differ within treatment ($P > 0.10$) for estrous cycling and prepubertal heifers (Table 9).

Table 8. Estrous response for estrous cycling and prepubertal heifers assigned to MGA-PG or 14-day CIDR-PG¹ treatment protocols (Mallory et al., 2010).

Item	MGA-PG	14-day CIDR-PG
Estrous response after PGF_{2α}		
Proportion	170/200	180/196
Percent	85 ^a	92 ^b
Estrous cycling²		
Proportion	135/154	138/151
Percent	88 ^x	91
Prepubertal³		
Proportion	35/46	42/45
Percent	76 ^{c,y}	93 ^d

^{a,b} Means within rows with different superscripts are different ($P = 0.01$).

^{c,d} Means within rows with different superscripts are different ($P = 0.03$).

^{x,y} Means within columns with different superscripts tend to differ ($P = 0.06$).

¹ See Figure 11 for a description of the treatment protocols.

² Estrous cycling = heifers assigned a RTS of 4 or 5.

³ Prepubertal = heifers assigned a RTS of 2 or 3.

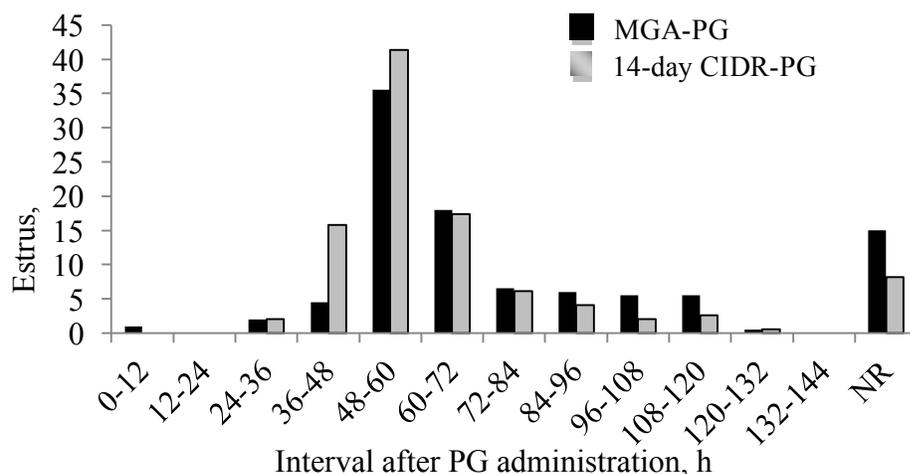


Figure 12. Percentage of heifers in MGA-PG and 14-day CIDR-PG treatments that exhibited estrus after PG: MGA-PG (black bar) and 14-day CIDR-PG (gray bar). NR = no estrous response.

Table 9. Mean and variance for interval from PG to estrus for estrous cycling and prepubertal heifers assigned to MGA-PG or 14-day CIDR-PG treatment protocols (See Figure 9 for a description of the treatment protocols; Mallory et al., 2010).

Item	MGA-PG	14-day CIDR-PG
Interval from PGF _{2α} to estrus, h (LS mean ± SE)	57.4 ± 2.5	56.2 ± 2.5
Estrous cycling	62.4 ± 2.4 ^{a,x}	55.4 ± 2.4 ^b
Prepubertal	52.4 ± 4.4 ^y	57.0 ± 4.4
Variance for interval to estrus after PGF _{2α}	466 ^c	282 ^d
Estrous cycling	432 ^c	272 ^d
Prepubertal	615 ^e	316 ^f

^{a,b} Means within rows with different superscripts are different (P = 0.04).

^{c,d} Variances within rows with different superscripts are different (P < 0.01).

^{e,f} Variances within rows with different superscripts are different (P < 0.05).

^{x,y} Means within columns with different superscripts are different (P = 0.04).

How do short- and long-term CIDR-based protocols compare in synchronizing ovulation prior to fixed-time artificial insemination in beef heifers?

Leitman et al. (2008) reported an improvement in synchrony of estrus and ovulation among CIDR Select treated heifers in comparison to Select Synch + CIDR treated contemporaries (Figure 13). There was more variance associated with the interval from PG to estrus (P<0.06) and ovulation (P<0.05) between prepubertal and estrous cycling heifers synchronized with the Select Synch + CIDR protocol compared to CIDR Select (Leitman et al., 2008). These data (Leitman et al., 2008) suggested that the CIDR Select protocol may facilitate FTAI more effectively in mixed groups of prepubertal and estrous cycling beef heifers compared with Select Synch + CIDR.

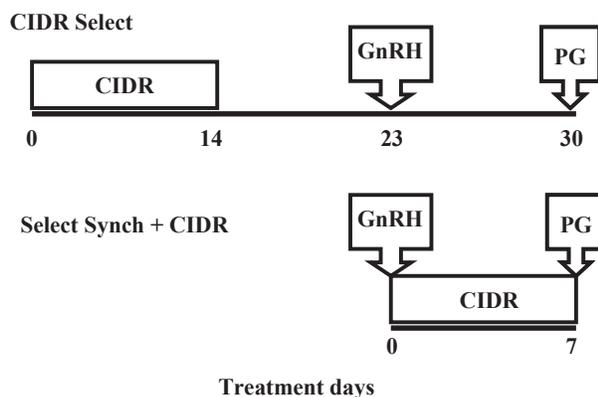


Figure 13. Comparison of CIDR Select and Select Synch + CIDR protocols (Leitman et al., 2008).

Busch et al. (2007) compared pregnancy rates resulting from fixed-time AI (FTAI) following administration of either one of two controlled internal drug release (CIDR)-based protocols (Figure 14). Heifers at three locations were assigned to one of two treatments within reproductive tract scores (RTS; 1 to 5, 1 =

immature, and 5 = cycling) by age and weight. Heifers assigned to CIDR Select received a CIDR insert from d 0 to 14 followed by GnRH 9 d after CIDR removal and PG 7 d after GnRH treatment. Heifers assigned to CO-Synch + CIDR were administered GnRH and received a CIDR insert, and PG and CIDR removal 7 d later (Figure 14).

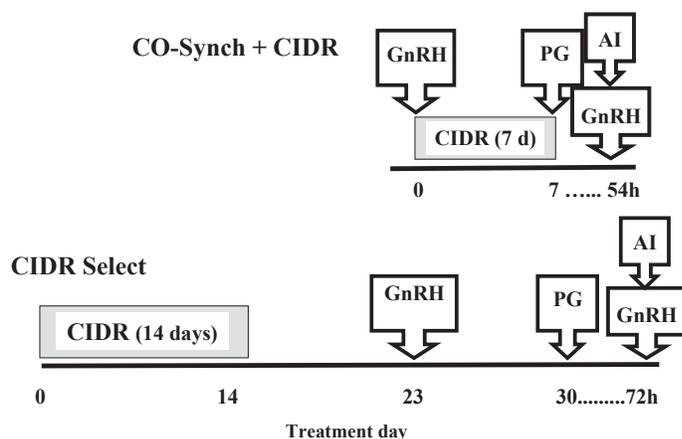


Figure 14. Treatment schedule for heifers assigned to the CIDR Select and CO-Synch + CIDR protocols (Busch et al., 2007).

Artificial insemination was performed at predetermined fixed-times for heifers in both treatments at 72 or 54 h after PG for the CIDR Select and CO-Synch + CIDR groups, respectively. All heifers were administered GnRH at the time of insemination. Pregnancy rates resulting from FTAI (Table 10) were significantly greater ($P = 0.02$) following the CIDR Select protocol (62%) compared to the CO-Synch + CIDR protocol (47%). In summary, the CIDR Select protocol resulted in a greater and more synchronous estrous response and significantly greater fixed-time AI pregnancy rates compared to the CO-Synch + CIDR protocol (Busch et al., 2007).

Table 10. Pregnancy rates of heifers in response to fixed-time AI and at the end of the breeding season (means \pm SE; Busch et al., 2007).

Item	Pregnancy rate to FTAI		Pregnancy rate at end of breeding season	
	Proportion	%	Proportion	%
CIDR Select	67/108	62 ^x	97/108	90
CO-Synch + CIDR	51/109	47 ^y	99/109	91

^{x,y} Means within a column with different superscripts are different, $P < 0.05$.

Development of the 14-day CIDR-PG protocol for heifers

Over the years, there have been questions raised regarding the utility of gonadotropin releasing hormone (GnRH) in estrus synchronization protocols for beef heifers (Wood-Follis et al., 2004; Lamb et al., 2006; Leitman et al., 2009a, b). Administration of GnRH at the beginning of an estrus synchronization protocol in beef heifers failed to demonstrate an increase in pregnancy rates resulting from fixed-time AI; however, the standard deviation of pregnancy rates was increased when GnRH was not included. These data suggested that incorporation of GnRH in a FTAI protocol may increase the uniformity of pregnancy rates in beef heifers across locations compared to protocols based on estrus detection alone (Lamb et al., 2006).

Leitman et al. (2009b; Figure 15) compared the CIDR Select and 14-day CIDR-PG protocols to determine the necessity of adding a GnRH injection for synchronization of estrus in beef heifers that were prepubertal or estrous cycling at the time treatments were initiated. Treatments were compared on the basis of estrous response and distribution of estrus after PG, and of synchronized AI conception and pregnancy rates.

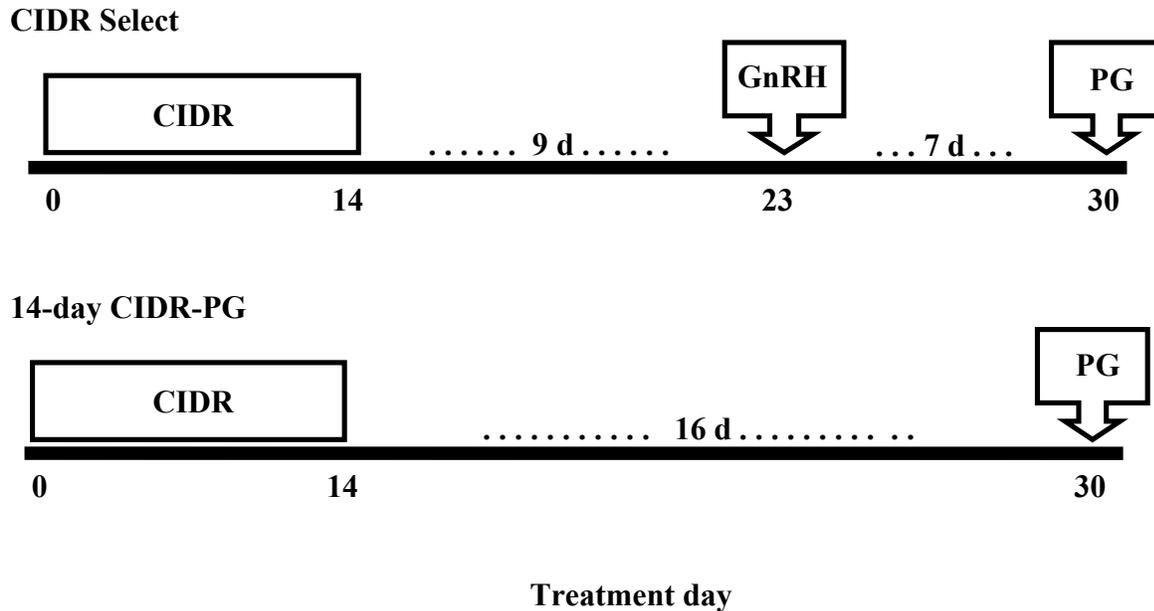


Figure 15. Treatment schedule for heifers assigned to the CIDR Select and 14-day CIDR-PG protocols. Heifers assigned to CIDR Select received a CIDR insert from d 0 to 14, GnRH on d 23, and PG on d 30. Heifers assigned to 14-day CIDR-PG received a CIDR insert from d 0 to 14, and PG on d 30 (Leitman et al., 2009b).

Figure 16 illustrates differences in estrous response after PG between the two treatments. In this experiment (Leitman et al., 2009b), differences in the variance for interval to estrus were detected based on the main effects of treatment and estrous cyclicity status as well as their interaction. Heifers assigned to the CIDR-PG protocol had a more highly synchronized estrus compared to heifers assigned to the CIDR Select protocol; and regardless of treatment, the prepubertal heifers had a more highly synchronized estrus compared to the estrous-cycling heifers. Improved synchrony of estrus observed among prepubertal heifers may be a result of a more highly synchronized estrous response following CIDR removal compared to estrous cycling heifers. Stage of cycle differences among estrous cycling heifers at CIDR insertion would perhaps explain the potential for reduced synchrony of estrus following CIDR removal compared to the prepubertal heifers. Both the estrous-cycling and prepubertal heifers assigned to the CIDR-PG protocol had a more highly synchronized estrus compared to their counterparts assigned to the CIDR Select protocol. While synchrony of estrus was similar between the estrous-cycling and prepubertal heifers assigned to the CIDR Select protocol, prepubertal heifers assigned to the CIDR-PG protocol had a more highly synchronized estrus compared to estrous-cycling heifers assigned to the CIDR-PG protocol.

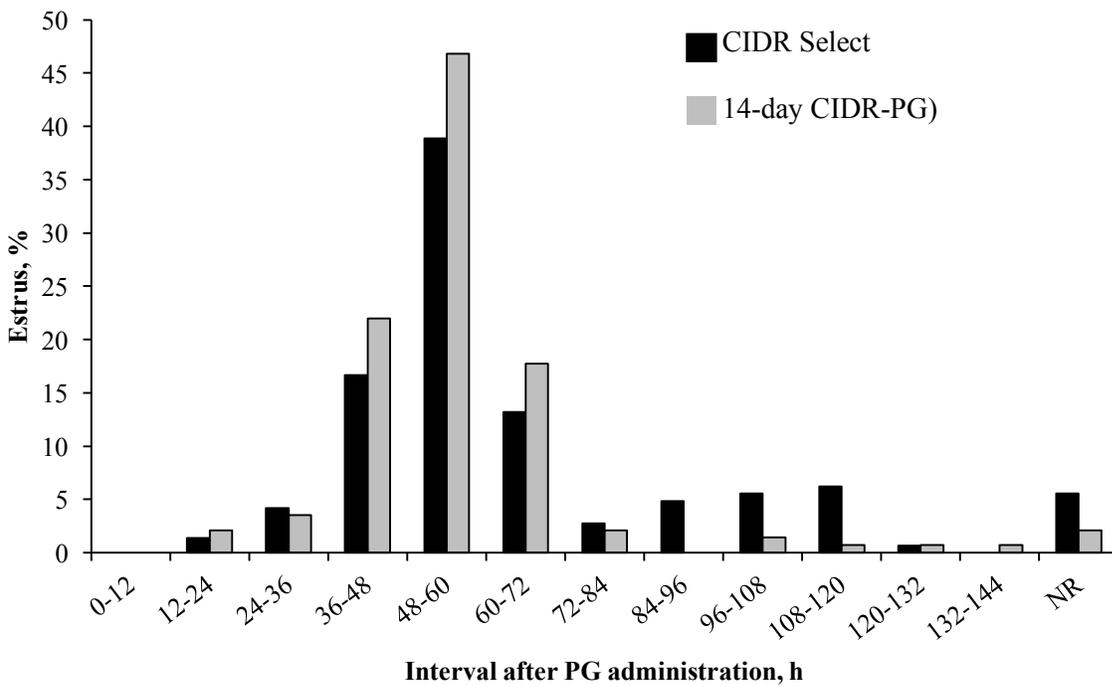


Figure 16. Percentage of heifers in the CIDR Select and 14-day CIDR-PG treatments that exhibited estrus after PG: CIDR Select (black bar) and 14-day CIDR-PG (gray bar). NR = no estrous response. See Figure 14 for a description of the treatment protocols (Leitman et al., 2009b).

We know from previous studies that there is no difference in estrous response following CIDR removal when comparing estrous-cycling or prepubertal heifers treated with a 14-d CIDR protocol (Leitman et al., 2008). Although the mean interval to estrus following CIDR removal was shorter for estrous-cycling heifers compared to prepubertal heifers, there was no difference in the variance for interval to estrus. Leitman et al. (2008) hypothesized that pre-synchronization with a progestin before GnRH and PG would be more effective in synchronizing estrus compared with 7-d CIDR-based or GnRH-PG estrus synchronization protocols. This hypothesis was tested and accepted. The study (Leitman et al., 2008) also revealed that estrous response and synchrony of estrus following removal of the CIDR after treatment for 14 days was similar between estrous cycling and prepubertal heifers. Additionally, over 88% of the heifers (estrous-cycling and prepubertal) were on d 7 or 8 of their estrous cycles 9 d following CIDR removal, coincident with the time at which GnRH was administered on d 23 of treatment of the CIDR Select protocol.

Arguably, given what we know regarding length of follicular waves (Savio et al., 1988; Sirois and Fortune, 1988), one might assume that a proportion of heifers may turn dominant follicles over on their own, prior to GnRH, independent of the need for GnRH to accomplish the same. Jaiswal et al. (2009) reported differences in 2-wave versus 3-wave patterns of ovarian follicular development in *Bos taurus* heifers. The prevalence of 2-wave versus 3-wave patterns was influenced by heifer age and/or maturity (Jaiswal et al., 2009). These authors (Jaiswal et al., 2009) suggest that more precise determination of predictive factors controlling patterns of follicular development in heifers will lead to the development of protocols that facilitate improvements in estrous cycle control and enhance opportunities to expand the use of FTAI.

These considerations may relate to the study by Leitman et al. (2009b), but fail to explain the significant improvement in synchrony of estrus for 14-day CIDR-PG compared to CIDR Select treated heifers. Although response to GnRH in heifers is reported to be inconsistent when compared to cows (Macmillan

and Thatcher, 1991; Pursley et al., 1995; Moreira et al., 2000), these data indicate that the addition of GnRH to a 14-d CIDR-PG protocol reduced the synchrony of estrus, despite similarities between treatments in estrous response. Schafer et al. (2006) and Leitman et al. (2009a) reported that the majority of heifers are on d 7 or 8 of the estrous cycle at the time GnRH is administered on d 23 of the CIDR Select protocol; therefore, the question arises as to the potential subsequent effect of administering GnRH to heifers at a point in their follicular wave at or during the time emergence of a new follicular wave begins. Given the fact that interval to estrus following PG was longer among CIDR Select versus 14-day CIDR-PG treated heifers, the effect of GnRH on subsequent follicular dynamics is in question.

Conception rate to AI (Table 11) tended to be greater for heifers assigned to 14-day CIDR-PG compared to CIDR Select, but was not influenced by estrous cyclicity status. Heifers assigned to 14-day CIDR-PG, however, had a higher pregnancy rate to AI compared to heifers assigned to CIDR Select. Pregnancy rate to AI (Table 11) was not influenced by estrous cyclicity status. These data point to the effectiveness of both protocols in inducing cyclicity in prepubertal heifers and successfully preparing heifers for breeding and subsequent pregnancy.

Perry et al. (2005; 2007) reported that use of protocols that control and/or manipulate follicular growth and development and increase the likelihood of ovulating optimal sized follicles may result in positive benefits on pregnancy rates in beef heifers. This is an important consideration based on studies that showed a relationship between ovulatory follicle size and pregnancy success in heifers (Perry et al., 2007) and cows (Vasconcelos et al., 2001; Lamb et al., 2001). Collectively, these reports support the concept that presynchronization is an effective means of manipulating follicle growth and development prior to a synchronized estrous period.

In summary from the experiment by Leitman et al. (2009b), similarities in estrous response following PG suggest that each of these long-term CIDR-based protocols was effective in synchronizing estrus in prepubertal and estrous-cycling beef heifers. The results from this experiment however, failed to confirm the hypothesis that the addition of GnRH on d 23 of the CIDR Select protocol results in a more highly synchronized estrus compared to 14-day CIDR-PG. Differences between treatments in the interval to estrus following PG, synchrony of estrus, and AI pregnancy rates during the synchronized period clearly suggested that further evaluation of these two CIDR-based protocols was required with and without the addition of GnRH and on the basis of estrous cyclicity status to determine the efficacy of these protocols for use in facilitating FTAI.

Table 11. Estrous response and interval to estrus after PGF_{2α} (PG), and AI conception rates and pregnancy rates for heifers assigned to controlled internal drug release (CIDR) Select or 14-day CIDR-PG¹ (Leitman et al., 2009a).

Item	CIDR Select	14-day CIDR-PG
Estrous response after PGF _{2α}		
Proportion	136/144	138/141
%	94	98
Interval from PGF _{2α} to estrus, h (LS mean ± SE)	61.5 ± 1.7 ^a	54.4 ± 1.7 ^b
Variance for interval to estrus after PGF _{2α}	508 ^a	262 ^b
Conception rate to AI		
Proportion	78/135 ^c	92/137 ^d
%	58	67
Pregnancy rate to AI		
Proportion	78/143 ^e	92/140 ^f
%	55	66
Pregnancy rate at the end of the breeding season		
Proportion	116/143	113/140
%	81	81

^{a,b} Means and/or variances within rows with different superscripts are different ($P \leq 0.01$).

^{c,d} Means within rows with different superscripts tend to differ ($P = 0.09$).

^{e,f} Means within rows with different superscripts are different ($P = 0.05$).

¹ See Figure 16 for a description of the treatment protocols.

How do the CIDR select (CIDR-GnRH-PG) and 14-day CIDR-PG protocols compare on the basis of pregnancy rates resulting from ftai in heifers?

Mallory et al. (2011) conducted an experiment to compare FTAI pregnancy rates after treatment with the CIDR Select and 14-day CIDR-PG treatment protocols (Figure 16). Pregnancy rates resulting from FTAI tended to differ between treatments with the advantage to heifers assigned to the 14-day CIDR-PG protocol (Table 12). Pretreatment estrous cyclicity status did not affect FTAI pregnancy rate; however, there was a trend toward higher FTAI pregnancy rates among estrous-cycling heifers assigned to the 14-day CIDR-PG protocol compared to those assigned to CIDR Select. No difference was detected between prepubertal heifers treated with CIDR Select or 14-day CIDR-PG protocols, possibly due to the low number of pre- and peripubertal heifers within each treatment.

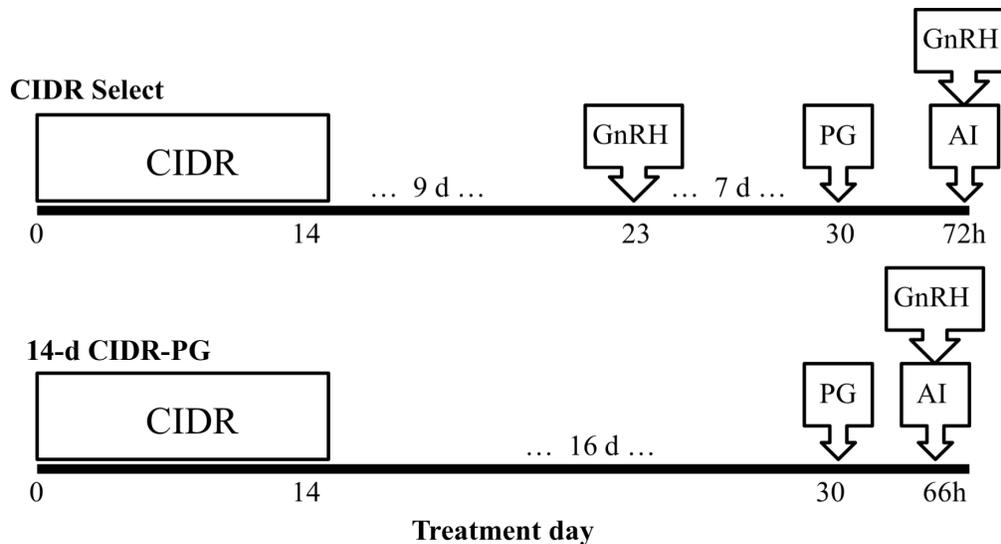


Figure 17. Treatment schedule for heifers assigned to the CIDR Select and 14-day CIDR-PG treatment protocols. Heifers assigned to the CIDR Select protocol received an EAZI-Breed CIDR insert from d 0 to d 14, GnRH on d 23, PG on d 30 followed by fixed-time AI at 72 h after PG administration. Heifers assigned to 14-day CIDR-PG received a CIDR insert from d 0 to 14 and PG on d 30 followed by fixed-time AI 66 h after PG administration (Mallory et al., 2011).

Timing of insemination for heifers assigned to the 14-day CIDR-PG protocol in this study was based on previous reports by Leitman et al. (2009a, b) and Mallory et al. (2010). Peak estrous response in those studies occurred 48 to 60 h after PG, and the peak AI date was 3 d after PG. Mean intervals to estrus after PG for the three experiments were 59.3 ± 2.8 h, 54.4 ± 1.7 h, and 56.2 ± 2.5 h, respectively (Leitman et al., 2009a, b; Mallory et al., 2010). Based on the consistency of these results (Mallory et al., 2011), timing of insemination at 66 h following the administration of PG was chosen. Timing of insemination after the CIDR Select protocol (72 h) was based on previous studies from our laboratory (Busch et al., 2007; Leitman et al., 2008).

In summary, the data clearly showed that the 14-day CIDR-PG protocol more effectively synchronized estrus prior to FTAI in heifers and provides an alternative to the CIDR Select protocol in facilitating expanded use of fixed-time artificial insemination (Mallory et al., 2011; 2013). This study further supports the results reported by Leitman et al. (2009b), indicating that GnRH is not required to successfully synchronize estrus prior to FTAI among heifers that are presynchronized with a 14-d CIDR treatment. Modification of the CIDR Select protocol to 14-day CIDR-PG allows producers to minimize trips through the chute and reduces cost associated with estrus synchronization and FTAI.

Table 12. AI pregnancy and final pregnancy rates for heifers assigned to CIDR Select or 14-day CIDR-PG treatment protocols (Mallory et al., 2011).

Item	CIDR Select	14-day CIDR-PG
Pregnancy rate to AI		
Proportion	98/192	124/200
Percent	51 ^a	62 ^b
Estrous cycling	83/158	102/162
Percent	53 ^c	63 ^d
Prepubertal	15/34	22/38
Percent	44	58
Pregnancy rate at the end of the breeding season		
Proportion	164/192	166/200
Percent	85	83
Estrous cycling	135/158	134/162
Percent	85	83
Prepubertal	29/34	32/38
Percent	85	84

^{a,b} Means within rows with different superscripts tended to differ (P = 0.07).

^{c,d} Means within rows with different superscripts tended to differ (P = 0.06).

How do the 14-day CIDR-PG and MGA-PG protocols compare on the basis of pregnancy rates resulting from FTAI in beef heifers?

Vraspir et al. (2013) recently compared pregnancy rates after fixed-time AI among Angus heifers (n=688) assigned to 14-day MGA-PG or CIDR-PG protocols. Heifers assigned the MGA-PG protocol were synchronized as illustrated in Figure 7 and inseminated by appointment at 72 hours after PG. Heifers in the 14-day CIDR-PG treatment were synchronized as illustrated in Figure 17 and inseminated at 66 hours after PG. Heifers in both treatments were administered GnRH at AI. Pregnancy rates after FTAI did not differ between treatments with 62% and 61% of the heifers conceiving to the MGA- and CIDR-treated groups, respectively. Vraspir et al. (2013) went on to report that 26% of the heifers in each treatment displayed a second estrus and were inseminated accordingly. In this case however there was a trend (P < .06) toward higher pregnancy rates among heifers assigned to the MGA- compared to the CIDR-treated groups (66 vs 56%, respectively). Overall pregnancy rates were similar between treatments (93% MGA-PG; 90% CIDR-PG).

Development of the 5-day CO-synch + CIDR and PG 6-day CIDR protocols

The 5-day CO-Synch + CIDR protocol with fixed-time AI at 60 hours after the first injection of PG and CIDR removal was recently added to the list of recommended protocols for fixed-time AI in beef heifers (Figure 18). This protocol was developed based on the hypothesis (Bridges et al., 2008) that reducing the length of CIDR treatment from 7 to 5 days in the CO-Synch + CIDR protocol would conceptually increase secretion of estradiol by the ovulatory follicle, decrease the incidence of induced ovulation of follicles with reduced estrogenic activity, and potentially result in improvements in FTAI pregnancy rates. The hypothesis was based on the premise that day 4 dominant follicles have higher intrafollicular concentrations of estradiol-17_β (E₂) and a greater ability to produce E₂ compared to older age follicles (Valdez et al., 2005). Bridges et al. (2009) reported that maximum preovulatory concentrations of estradiol tended to be greater in 5-day compared to 7-day CIDR-treated cows that failed to respond to GnRH at CIDR insertion, and that postovulatory circulating concentrations of progesterone were greater among 5-day compared to the

7-day treated cows. Increased follicular concentrations of E₂ and elevated postovulatory concentrations of progesterone are believed to reflect greater physiological maturity of the dominant follicle and to result in higher pregnancy rates resulting from AI (Lopez et al., 2005; Perry et al., 2005). Therefore, Bridges et al. (2008) proposed that if CIDR removal and AI are more accurately timed with the 5-day protocol to coincide with follicular development, higher AI pregnancy rates may be achieved.

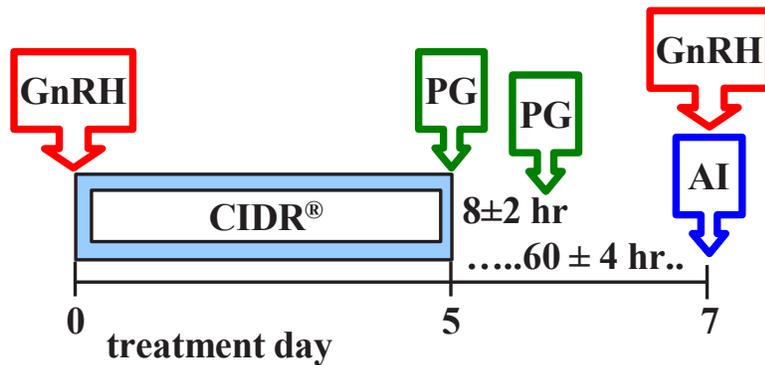


Figure 18. The 5-day CO-Synch + CIDR protocol for fixed-time AI in beef heifers.

Grant et al. (2011) and Perry et al. (2012) designed experiments (Figure 19) to determine whether controlling follicular development to optimize follicle size and estradiol exposure would be effective in maximizing pregnancy success. The objectives of the studies (Perry et al., 2012) were to determine: 1) if inducing luteal regression 3 days prior to an injection of GnRH improved control of follicular turnover, and 2) if inducing luteal regression 3 days before a modified CIDR protocol improved pregnancy success following fixed-time AI.

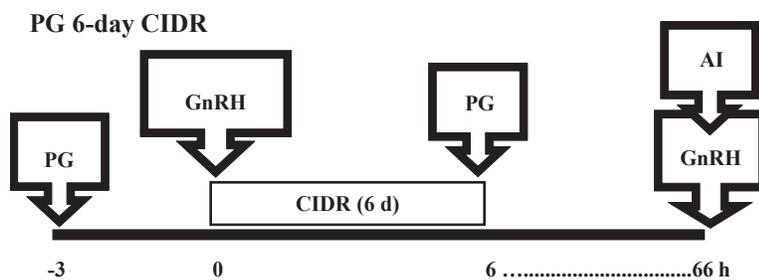


Figure 19. The PG 6-day CIDR protocol.

Regrettably there are no published studies illustrating estrus distribution patterns for the 5-day CO-Synch + CIDR or PG 6-day CIDR protocols in beef heifers on the basis of estrous cyclicity status of heifers at the time treatments are imposed. However, recently Bridges et al. (2014) published results from a study comparing the 5-day CO-Synch + CIDR, PG 6-day CIDR, and 14-day CIDR-PG protocols. Pregnancy rates resulting from FTAI were compared among 12-15 month old heifers on ranches in four states. Although pretreatment estrous cyclicity status was determined for heifers assigned to each of these treatments, given the differences in length of the respective treatments it would have been interesting to compare results had the heifers been assigned to treatment on the basis of cyclicity at the initiation of the 14-day CIDR treatment. Instead, heifers were pre-assigned to treatment, and then assessed for pubertal status

at the respective times treatments were imposed. Fixed-time AI pregnancy success did not differ among treatments; however as reported from previous studies, heifers that reached puberty prior to treatment initiation had greater pregnancy success compared with heifers that were prepubertal.

Recently, Kasimanickam et al. (2015) compared pregnancy rates of heifers after FTAI that were synchronized with the 5-day CO-Synch + CIDR or the 14-day CIDR-PG protocol. There were no differences between treatments in pregnancy rates resulting from FTAI (5-day CO-Synch + CIDR, 56%; 14-day CIDR-PG, 55%) however heifers assigned to the 14-day CIDR-PG protocol were inseminated 72 hours after PG instead of the recommended 66 hours.

Split-time AI: Delayed insemination of non-estrous beef heifers in a timed artificial insemination protocol

Thomas et al. (2014a,b) tested the hypothesis that pregnancy rates in beef heifers after FTAI may be improved by delaying insemination of heifers that have not expressed estrus prior to the standard FTAI time. Split-time artificial insemination (STAI) following administration of the 14-d CIDR-PG protocol allows heifers to be managed based on estrous response at the time of insemination, which facilitates an increase in pregnancy rates compared to fixed-time artificial insemination (FTAI). In STAI, insemination is delayed for non-estrous females by 20 to 24 h, whereas in FTAI, all heifers are inseminated at a single time. All heifers were administered GnRH at 66 h irrespective of estrous status in the original field trials that compared ST- and FTAI. The working hypothesis in that study (Thomas et al., 2014a,b) was that delayed insemination of non-estrous heifers would better align the timing of insemination with the timing of GnRH-induced ovulations. This approach to breeding management increased overall pregnancy rates in beef heifers.

Later experiments (Bishop et al., 2016a,b) evaluated the optimal timing of GnRH administration when using STAI. Results from these studies clearly demonstrated that administration of GnRH was not required for heifers that exhibit estrus prior to AI, and that administration of GnRH to heifers that fail to exhibit estrus by 66 h could be delayed to 90 h, concurrent with insemination. This work confirmed that higher pregnancy rates resulting from STAI compared to FTAI are due primarily to higher estrous response rates prior to insemination, in contrast to the theory that timing of insemination is more optimally aligned relative to GnRH-induced ovulations (Thomas et al., 2014a,b). Despite the high overall estrous response that is observed in heifers when split-time AI is practiced, a percentage of heifers fail to exhibit estrus prior to 90 h after PG (Bishop et al., 2016a,b). Pregnancy rates resulting from AI are generally reduced among these heifers compared with those that exhibit estrus (Mallory et al., 2013; Perry and Smith, 2015), despite the fact GnRH is routinely administered concurrent with AI. Bishop et al. (2016b) reported that when split-time AI was used in conjunction with the 14-day CIDR-PG protocol comparable pregnancy rates were achieved among non-estrous heifers at 90 hours after PG regardless of whether or not GnRH was administered concurrent with AI. These results (Bishop et al., 2016b) continue to raise questions regarding the general efficacy of GnRH in beef heifers.

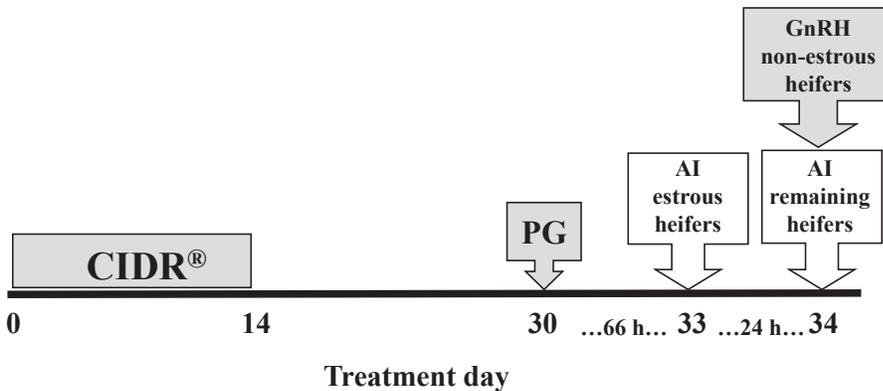


Figure 20. The 14-day CIDR-PG protocol for heifers with split-time AI (Bishop et al., 2016b; Thomas et al., 2016).

Using estrus detection aids to facilitate split-time AI.

To identify animals that have not expressed estrus, EstroTECT® estrus detection aids can be applied at the time of the final PG administration. EstroTECT® patches are designed with a scratch-off coating that is removed progressively as an estrous animal stands to be mounted. Because of this design, producers can have a high degree of confidence about the estrus status of females. When using split-time AI, an animal is considered as having expressed estrus when over 50% of the scratch-off coating has been removed from the EstroTECT® patch (Thomas et al., 2016).

Comparing pregnancy rates resulting from fixed-time AI on the basis of reproductive tract score

In 1996 extension specialists, veterinarians, beef producers and allied industry in Missouri linked arms to develop and implement a comprehensive plan that would impact long-term sustainability of beef herds across the state. The plan was focused on the cyclical reproductive processes involved with cow-calf operations and involves five basic steps. These five steps have built equity in herds that embraced the plan, and 20 years later the Missouri Show-Me-Select Replacement Heifer Program™ has impacted the cattle industry state wide. The program incorporates all available tools to support long-term health, reproduction, and genetic improvement of replacement beef heifers and includes provisions for ownership; health and vaccination schedules; parasite control; implant use; weight, pelvic measurement and reproductive tract score; estrous synchronization and artificial insemination; service-sire requirements for BW- or CE-EPD; early pregnancy diagnosis, fetal aging, fetal sexing, and body condition score. These steps include: 1) Create an understanding of the importance of heifer development based on reproductive outcomes; 2) Implement changes in heifer development that will eventually spill over into the cow herd; 3) Emphasize the importance of reproductive management which becomes apparent as changes are implemented; 4) Expand producer focus to genetic improvement; 5) Emphasize to participating herds that creation of a value-added product requires a re-evaluation of marketing strategies.

The Missouri Show-Me-Select Replacement Heifer Program™ provides a unique opportunity to collect reproductive data on large numbers of beef heifers from the perspective of pretreatment estrous cyclicity status, the respective protocols to synchronize estrus, and pregnancy rates resulting from FTAI. Prebreeding exams [reproductive tract scores (RTS)] are performed by veterinarians on all heifers enrolled in the program prior to each breeding season. Additionally, pregnancy diagnoses are performed within 90 days from the start of the breeding period and re-confirmed after the end of the breeding season. In recent years, program participants have increased the use of fixed-time AI programs in their herds. Field data collected

from 2010-2015 were used to evaluate relationships between RTS and pregnancy outcome after FTAI. As previously reviewed, the reproductive tract scoring system ranges from 1 to 5 (Anderson et al., 1991): 1=infantile; 2 and 3=noncycling/prepubertal; 4 and 5=cycling/pubertal. A summary of RTS and FTAI pregnancy rate for 23,481 heifers that were evaluated from 2010-2015 is provided in Table 14 (Locke et al., 2016).

Table 14. Pregnancy rates resulting from fixed-time AI based on reproductive tract score (FTAI; Fall 2010 – Fall 2015). Missouri Show-Me-Select Replacement Heifer Program™* (Locke et al., 2016).

Reproductive Tract Score (RTS)	1	2	3	4	5
Number of heifers	136	768	6,634	7,803	8,140
Number pregnant	8	221	3,099	3,915	4,242
FTAI pregnancy rate (%)	6%	29%	47%	50%	52%

*Pregnancy rates resulting from fixed-time AI based on RTS. These data include pregnancy rates for 23,481 heifers that were inseminated beginning during the fall of 2010 through fall of 2015.

These data support the practice of establishing prebreeding criteria for identification of heifers that are good candidates for a FTAI program. The data are further summarized in Table 15 to evaluate three of the recommended protocols to facilitate FTAI on the basis of estrous cyclicity status of heifers prior to the time treatments to synchronize estrus were initiated. Data for heifers assigned a RTS of 1 were excluded from this summary. These data indicate that evaluation of reproductive status of heifers prior to the first breeding season is useful in determining success of the development period and in determining which protocol to use to synchronize estrus prior to FTAI.

Table 15. Pregnancy rates after FTAI based on reproductive tract score and protocol used to synchronize estrus. Missouri Show-Me-Select Replacement Heifer Program™* (Locke et al., 2016).

Protocol	Reproductive tract score (RTS)			
	Non-cycling (2 and 3)		Cycling (4 and 5)	
7-day CO-Synch + CIDR	121/333	36% ^{a,x}	305/689	44% ^{b,x}
MGA-PG	81/230	35% ^{a,x}	265/564	47% ^{b,x}
14-day CIDR-PG	3,118/6,839	46% ^{a,y}	7,583/14,684	52% ^{b,y}

^{a,b} Percentages within rows with different superscripts differ ($P < 0.01$).

^{x,y} Percentages within columns with different superscripts differ ($P < 0.01$).

The Missouri Show-Me-Select Replacement Heifer Program™ continues to see an increase in the overall percentage of heifers exposed for AI, and the majority of these heifers are being inseminated at pre-determined fixed times. An important consideration related to the success of these AI programs and the expanded use of AI in Missouri is the fact that all heifers are required to undergo a prebreeding examination to determine estrous cyclicity status. This practice allows for better determination of the appropriate estrus synchronization protocol from which to choose, but at the same time provides critical information that is supportive in situations where troubleshooting is required.

These conclusions are supported by a study reported recently by Gutierrez et al. (2014) which involved a comparison of heifers that were inseminated on the basis of FTAI followed by a natural service clean-up period versus natural service only. Gutierrez et al. (2014) reported that RTS influenced both the number of beef heifers that became pregnant during the breeding season and the time at which heifers became pregnant.

Summary and conclusions

Protocols for inducing and synchronizing a fertile estrus in replacement beef heifers in which progestins are used provide opportunities for beef producers to synchronize estrus and ovulation and facilitate FTAI. These data suggest that methods of inducing and synchronizing estrus for replacement beef heifers provide the opportunity to significantly expand the use of AI in the U.S. cowherd.

References

- Anderson K. J., D. G. LeFever, J. S. Brinks, and K. G. Odde. 1991. The use of reproductive tract scoring in beef heifers. *Agri-Practice* 1991; 12:106-111.
- Anderson, L. H., C. M. McDowell, and M. L. Day. 1996. Progesterin-induced puberty and secretion of luteinizing hormone in heifers. *Biol. Reprod.* 54:1025-1031.
- Atkins, J. A., D. C. Busch, J. F. Bader, D. H. Keisler, D. J. Patterson, M. C. Lucy, and M. F. Smith. 2008. Gonadotropin-releasing hormone-induced ovulation and luteinizing hormone release in beef heifers: Effect of day of the cycle. *J. Anim. Sci.* 86:83-93.
- Berardinelli, J. G., R. A. Dailey, R. L. Butcher, and E. K. Inskeep. 1979. Source of progesterone prior to puberty in beef heifers. *J. Anim. Sci.* 49:1276-1281.
- Bishop, B. E., J. M. Thomas, J. M. Abel, S. E. Pooch, M. R. Ellersieck, M. F. Smith, and D. J. Patterson. 2016a. Split-time artificial insemination in beef cattle: I. Using estrous response to determine optimal time(s) at which to administer GnRH in beef heifers and postpartum cows. *Theriogenology*. Volume 86, Number 4: 1102-1110.
- Bishop, B. E., J. M. Thomas, J. M. Abel, S. E. Pooch, M. R. Ellersieck, M. F. Smith, and D. J. Patterson. 2016b. Split-time artificial insemination in beef cattle. II. Comparing pregnancy rates among non-estrous heifers based on administration of GnRH at AI. *Theriogenology*. In press.
- Bridges, G. A., L. A. Helser, D. E. Gru, M. L. Mussard, C. L. Gasser, and M. L. Day. 2008. Decreasing the interval between GnRH and PGF_{2α} from 7 to 5 days and lengthening proestrus increased timed-AI pregnancy rates in beef cows. *Theriogenology* 69:843-851.
- Bridges, G. A., M. L. Mussard, L. A. Hesler, and M. L. Day. 2009. Comparison of follicular dynamics and hormone concentrations between the 7 d and 5 d CO-Synch + CIDR program in two-year-old beef cows. *J. Anim. Sci.* 87(E-Suppl. 2):372. (Abstr.)
- Bridges, G. A., S. L. Lake, S. G. Kruse, S. L. Bird, B. J. Funnell, R. Arias, J. A. Walker, J. K. Grant, and G. A. Perry. 2014. Comparison of three CIDR-based fixed-time AI protocols in beef heifers. *J. Anim. Sci.* 92:3127-3133.
- Brown, L. N., K. G. Odde, D. G. LeFever, M. E. King, and C. J. Neubauer. 1988. Comparison of MGA-PGF_{2α} to Syncro-Mate B for estrous synchronization in beef heifers. *Theriogenology* 30:1.
- Burfening, P. J. 1979. Induction of puberty and subsequent reproductive performance. *Theriogenology* 12:215-221.
- Busch, D. C., D. J. Wilson, D. J. Schafer, N. R. Leitman, J. K. Haden, M. R. Ellersieck, M. F. Smith, and D. J. Patterson. 2007. Comparison of CIDR-based estrus synchronization protocols prior to fixed-time AI on pregnancy rate in beef heifers. *J. Anim. Sci.* 85:1933-1939.
- Deutscher, G. H. 2000. Extending interval from seventeen to nineteen days in the melengestrol acetate-prostaglandin estrous synchronization program for heifers. *Prof. Anim. Sci.* 16:164-168.
- Dziuk, P. J., and R. A. Bellows. 1983. Management of reproduction in beef cattle, sheep and pigs. *J. Anim. Sci.* 57(Suppl.2), 355.

- Federal Register. March 26, 1997. New animal drugs for use in animal feeds; Melengestrol Acetate. Vol. 62. No.58. pp.14304-14305.
- Garcia-Winder, M., P. E. Lewis, D. R. Deaver, V. G. Smith, G. S. Lewis, and E. K. Inskeep. 1986. Endocrine profiles associated with the life span of induced corpora lutea in postpartum beef cows. *J. Anim. Sci.* 62:1353-1362.
- Gonzalez-Padilla, E., R. Ruiz, D. LeFever, A. Denham, and J. N. Wiltbank. 1975. Puberty in beef heifers. III. Induction of fertile estrus. *J. Anim. Sci.* 40:1110-1118.
- Grant, J. K., F. M. Abreu, N. L. Hojer, S. D. Fields, B. L. Perry, and G. A. Perry. 2011. Influence of inducing luteal regression before a modified controlled internal drug-release device treatment on control of follicular development. *J. Anim. Sci.* 89:3531-3541.
- Gutierrez, K., R. Kasimanickam, A. Tibary, J. G. Gay, J. P. Kastelic, J. B. Hall, and W. D. Whittier. 2014. Effect of reproductive tract scoring on reproductive efficiency in beef heifers bred by timed artificial insemination and natural service versus only natural service. *Theriogenology*. 81:7; 918-924.
- Hall, J. B., R. B. Staigmiller, R. E. Short, R. A. Bellows, M. D. MacNeil, and S. E. Bellows. 1997. Effect of age and pattern of gain on induction of puberty with a progestin in beef heifers. *J. Anim. Sci.* 75:1606-1611.
- Henricks, D. M., J. R. Hill, and J. F. Dickey. 1973. Plasma ovarian hormone levels and fertility in beef heifers treated with melengestrol acetate (MGA). *J. Anim. Sci.* 37:1169-1175.
- Imwalle, D. B., D. J. Patterson, and K. K. Schillo. 1998. Effects of melengestrol acetate on onset of puberty, follicular growth, and patterns of luteinizing hormone secretion in beef heifers. *Biol. Reprod.* 58:1432-1436.
- Imwalle, D. B., D. L. Fernandez, and K. K. Schillo. 2002. Melengestrol acetate blocks the preovulatory surge of luteinizing hormone, the expression of behavioral estrus and ovulation in beef heifers. *J. Anim. Sci.* 80:1280-1284.
- Jaiswal, R. S., J. Singh, L. Marshall, and G. P. Adams. 2009. Repeatability of 2-wave and 3-wave patterns of ovarian follicular development during the bovine estrous cycle. *Theriogenology* 72: 81-90.
- Kasimanickam, R., S. Schroeder, J. B. Hall, and W. D. Whittier. 2015. Fertility after implementation of long- and short-term progesterone-based ovulation synchronization protocols for fixed-time artificial insemination in beef heifers. *Theriogenology* 83:1226-1232.
- Kojima, F. N., J. F. Bader, J. E. Stegner, D. J. Schafer, J. C. Clement, R. L. Eakins, M. F. Smith, and D. J. Patterson. 2004. Substituting EAZI-BREED CIDR inserts (CIDR) for melengestrol acetate (MGA) in the MGA Select protocol in beef heifers. *J. Anim. Sci.* 82(Suppl. 1):255.
- Lamb, G. C., D. W. Nix, J. S. Stevenson, and L. R. Corah. 2000. Prolonging the MGA-prostaglandin $F_{2\alpha}$ interval from 17 to 19 days in an estrus synchronization system for heifers. *Theriogenology* 53:691-698.
- Lamb, G. C., J. S. Stevenson, D. J. Kesler, H. A. Garverick, D. R. Brown, and B. E. Salfen. 2001. Inclusion of an intravaginal progesterone insert plus GnRH and prostaglandin $F_{2\alpha}$ for ovulation control in postpartum suckled beef cows. *J. Anim. Sci.* 79:2253-2259.
- Lamb, G. C., J. E. Larson, T. W. Geary, J. S. Stevenson, S. K. Johnson, M. L. Day, R. P. Ansotegui, D. J. Kesler, J. M. DeJarnette, and D. G. Landblom. 2006. Synchronization of estrus and artificial insemination in replacement beef heifers using gonadotropin-releasing hormone, prostaglandin $F_{2\alpha}$ and progesterone. *J. Anim. Sci.* 84:3000-3009.

- Leitman, N. R., D. C., Busch, J. F. Bader, D. A. Mallory, D. J. Wilson, M. C. Lucy, M. R. Ellersieck, M. F. Smith, and D. J. Patterson. 2008. Comparison of protocols to synchronize estrus and ovulation in estrous cycling and prepubertal beef heifers. *J. Anim. Sci.* 86:1808-1818.
- Leitman, N. R., D. C. Busch, D. A. Mallory, D. J. Wilson, M. R. Ellersieck, M. F. Smith, and D. J. Patterson. 2009a. Comparison of long-term CIDR-based protocols to synchronize estrus in beef heifers. *Animal Reproduction Science* 114: 345-355.
- Leitman, N. R., D. C. Busch, D. J. Wilson, D. A. Mallory, M. R. Ellersieck, M. F. Smith, and D. J. Patterson. 2009b. Comparison of controlled internal drug release insert-based protocols to synchronize estrus in prepubertal and estrous-cycling beef heifers. *J. Anim. Sci.* 87: 3976-3982.
- Locke, J. W. C., J. M. Thomas, B. E. Bishop, J. M. Abel, S. E. Pooock, D. S. Brown, J. E. Decker, and D. J. Patterson. 2016. The Show-Me-Select Replacement Heifer Program: Adding value to beef herds in Missouri. *J. Anim. Sci.* In press.
- Lopez, H., R. Sartori, and M. C. Wiltbank. 2005. Reproductive hormones and follicular growth during development of one or multiple dominant follicles in cattle. *Biol. Reprod.* 72:788-795.
- Lucy, M. C., H. J. Billings, W. R. Butler, L. R. Ehnis, M. J. Fields, D. J. Kesler, J. E. Kinder, R. C. Mattos, R. E. Short, W. W. Thatcher, R. P. Wettemann, J. V. Yelich, and H. D. Hafs. 2001. Efficacy of an intravaginal progesterone insert and an injection of PG F_{2α} for synchronizing estrus and shortening the interval to pregnancy in postpartum beef cows, peripubertal beef heifers, and dairy heifers. *J. Anim. Sci.* 79:982-995.
- Macmillan, K. L., and W. W. Thatcher. 1991. Effects of an agonist on gonadotropin-releasing hormone on ovarian follicles in cattle. *Biol. Reprod.* 45: 883-889.
- Macmillan, K. L., and A. J. Peterson. 1993. A new intravaginal progesterone releasing device for cattle (CIDR-B) for oestrous synchronization, increasing pregnancy rates and the treatment of post-partum anoestrus. *Anim. Reprod. Sci.* 33:1-25.
- Mallory, D. A., D. J. Wilson, D. C. Busch, M. R. Ellersieck, M. F. Smith, and D. J. Patterson. 2010. Comparison of long-term progestin-based estrus synchronization protocols in beef heifers. *J. Anim. Sci.* 88:3568-3578.
- Mallory, D. A., J. M. Nash, M. R. Ellersieck, M. F. Smith, and D. J. Patterson. 2011. Comparison of long-term progestin-based protocols to synchronize estrus before fixed-time artificial insemination in beef heifers. *J. Anim. Sci.* 89:1358-1365.
- Mallory, D. A., S. L. Lock, D.C. Woods, S. E. Pooock, and D. J. Patterson. 2013. Comparison of sex-sorted and conventional semen within a fixed-time artificial insemination protocol for dairy heifers. *J. Dairy. Sci.* 96:854-856.
- Moreira, F., R. L. de la Sota, T. Diaz, and W. W. Thatcher. 2000. Effect of day of the estrous cycle at the initiation of a timed artificial insemination protocol on reproductive responses in dairy heifers. *J. Anim. Sci.* 78: 1568-1576.
- Patterson, D. J., G. H. Kiracofe, J. S. Stevenson, and L. R. Corah. 1989. Control of the bovine estrous cycle with melengesrol acetate (MGA): A review. *J. Anim. Sci.* 67:1895-1906.
- Patterson, D. J., L. R. Corah, and J. R. Brethour. 1990. Response of prepubertal *Bos taurus* and *Bos indicus* x *Bos taurus* heifers to melengestrol acetate with or without gonadotropin-releasing hormone. *Theriogenology* 33:661-669.
- Patterson, D. J., J. M. Kearnan, N. W. Bradley, K. K. Schillo, and B. L. Woods. 1993. Estrus response and fertility in yearling beef heifers after chronic treatment with an oral progestogen followed by prostaglandin F_{2α}. University of Kentucky Beef Cattle Research Report. Progress Report 353. Pp. 31-33.

- Patterson, D. J., S. L. Wood, and R. F. Randle. 2000a. Procedures that support reproductive management of replacement beef heifers. *Proc. Am.Soc. Anim. Sci.*, 1999. Available at: <http://www.asas.org/jas/symposia/proceedings/0902.pdf>. Accessed August 3, 2000.
- Patterson, D. J., S. L. Wood, F. N. Kojima, and M. F. Smith. 2000b. Current and emerging methods to synchronize estrus with melengestrol acetate. In: 49th Annual Beef Cattle Short Course Proceedings “Biotechnologies of Reproductive Biology”. Pp. 45-66. University of Florida, Gainesville.
- Patterson, D. J., F. N. Kojima, and M. F. Smith. 2003. A review of methods to synchronize estrus in replacement heifers and postpartum beef cows. *J. Anim. Sci.* 81(E. Suppl. 2):E166-E177. Online. Available: <http://www.asas.org/symposia/03esupp2/jas2402.pdf>. Accessed June 19, 2003.
- Patterson, D. J., D. J. Schafer, D. C. Busch, N. R. Leitman, D. J. Wilson, and M. F. Smith. 2006. Review of estrus synchronization systems: MGA. In: Proceedings Applied Reproductive Strategies in Beef Cattle. St. Joseph, MO. Pp. 63-103.
- Perry, G.A. and M.F. Smith. 2015. Management factors that impact the efficiency of applied reproductive strategies. In: Proceedings, Applied Reproductive Strategies in Beef Cattle. August 17-18. Davis, CA. pp. 208–232.
- Perry, G.A., M.F. Smith, M.C. Lucy, J.A. Green, T.E. Parks, M.D. MacNeil, A.J. Roberts, and T.W. Geary. 2005. Relationship between follicle size at insemination and pregnancy success. *Proc. Natl. Acad. Sci. USA* 102:5268-5273.
- Perry, G. A., M. F. Smith, A. J. Roberts, M. D. MacNeil, and T. W. Geary. 2007. Relationship between size of the ovulatory follicle and pregnancy success in beef heifers. *J. Anim. Sci.* 85:684-689.
- Perry, G. A., J. K. Grant, J. A. Walker, G. A. Bridges, S. G. Kruse, S. Bird, K. Heaton, R. Arias, and S. L. Lake. 2012. Comparison of three CIDR-based fixed-time AI protocols for beef heifers. *J. Anim. Sci.* 90 (Suppl. 3):237.
- Prybil, M. K., and W. R. Butler. 1978. The relationship between progesterone secretion and the initiation of ovulation in postpartum beef cows. *J. Anim. Sci.* 47(Suppl. 1):383.
- Pursley, J. R., M. O. Mee, and M. C. Wiltbank. 1995. Synchronization of ovulation in dairy cows using PGF_{2α} and GnRH. *Theriogenology* 44:915-924.
- Rawlings, N. C., L. Weir, B. Todd, J. Manns, and J. Hyland. 1980. Some endocrine changes associated with the postpartum period of the suckling beef cow. *J. Reprod. Fertil.* 60:301-308.
- Savio, J. D., L. Kennan, M. P. Boland, and J. F. Roche. 1988. Pattern of growth of dominant follicles during the oestrus cycle in heifers. *J. Reprod. Fertil.* 83:663-671.
- Schafer, D. J., D. C. Busch, M. F. Smith, and D. J. Patterson. 2006. Characterization of follicular dynamics, timing of estrus, and response to GnRH and PG in replacement beef heifers after presynchronization with a 14-day CIDR. *J. Anim. Sci.* 84(Suppl. 1):49.
- Schafer, D. W., J. S. Brinks, and D. G. LeFever. 1990. Increased calf weaning weight and weight via estrus synchronization. Beef Program Report. Colorado State University. pp. 115-124.
- Seidel, G. E. Jr. 1995. Reproductive biotechnologies for profitable beef production. *Proc. Beef Improvement Federation*. Sheridan, WY. Pp. 28-39.
- Sheffel, C. E., B. R. Pratt, W. L. Ferrell, and E. K. Inskeep. 1982. Induced corpora lutea in the postpartum beef cow. II. Effects of treatment with progestogen and gonadotropins. *J. Anim. Sci.* 54:830-836.
- Sirois, J., and J. E. Fortune. 1988. Ovarian follicular dynamics during the estrous cycle in heifers monitored by real-time ultrasonography. *Biol. Reprod.* 39:308-317.

- Smith, R. K., and M. L. Day. 1990. Mechanism of induction of puberty in beef heifers with melengestrol acetate. In: Ohio Beef Cattle Res. and Ind. Rep. pp 137-142. Columbus, OH.
- Tauck, S. A., J. R. C. Wilkinson, J. R. Olsen, J. N. Janitell, and J. G. Berardinelli. 2007. Comparison of controlled internal drug release device and melengestrol acetate as progestin sources in an estrous synchronization protocol for beef heifers. *Theriogenology* 68:162-167.
- Thomas, J. M., S. L. Lock, S. E. Pooock, M. R. Ellersieck, M. F. Smith, and D. J. Patterson. 2014a. Delayed insemination of non-estrous cows improves pregnancy rates when using sex-sorted semen in timed artificial insemination of suckled beef cows. *J. Anim. Sci.* 92:1745-1750.
- Thomas, J. M., S. E. Pooock, M. R. Ellersieck, M. F. Smith, and D. J. Patterson. 2014b. Delayed insemination of non-estrous heifers and cows when using conventional semen in timed artificial insemination. *J. Anim. Sci.* 92:4189-4197.
- Thomas, J. M., B. E. Bishop, J. M. Abel, J. W. C. Locke, S. E. Pooock, D. S. Brown, M. F. Smith, and D. J. Patterson. 2016. Split-time AI: Using estrus detection aids to optimize timed artificial insemination. University of Missouri Extension MP739.
- Utter, S. D., and L. R. Corah. 1994. Influence of dietary energy levels on reproductive function and fertility in yearling beef heifers. Kansas State University Cattlemen's Day Proceedings. Pp. 104-106.
- Valdez, K. E., S. P. Cuneo, and A. M. Turzillo. 2005. Regulation of apoptosis in the atresia of dominant bovine follicles of the first follicular wave following ovulation. *Reproduction* 130:71-81.
- Vasconcelos, J. L., R. Sartori, H. N. Oliveira, J. G. Guenther, and M. C. Wiltbank. 2001. Reduction in size of the ovulatory follicle reduces subsequent luteal size and pregnancy rate. *Theriogenology* 56:300-314.
- Vraspir, R. A., A. F. Summers, D. O'Hare, L.D. Rowden, and R. N. Funston. 2013. Comparison of melengestrol acetate and controlled internal drug-release long-term progestin-based synchronization protocols on fixed-time artificial-insemination pregnancy rate in beef heifers. *Professional Animal Scientist* 29:575-579.
- Wetteman, R. P., and H. D. Hafs. 1973. Pituitary and gonadal hormones associated with fertile and nonfertile inseminations at synchronized and control estrus. *J. Anim. Sci.* 36:716-721.
- Wood, S. L., M. C. Lucy, M. F. Smith, and D. J. Patterson. 2001. Improved synchrony of estrus and ovulation with addition of GnRH to a melengestrol acetate-prostaglandin $F_{2\alpha}$ estrus synchronization treatment in beef heifers. *J. Anim. Sci.* 79:2210-2216.
- Wood-Follis, S. L., F. N. Kojima, M. C. Lucy, M. F. Smith, and D. J. Patterson. 2004. Estrus synchronization in beef heifers with progestin-based protocols. I. Differences in response based on pubertal status at the initiation of treatment. *Theriogenology* 62:1518-1528.
- Zimbelman, R. G. 1963. Maintenance of pregnancy in heifers with oral progestogens. *J. Anim. Sci.* 22:868.
- Zimbelman, R. G., and L. W. Smith. 1966. Control of ovulation in cattle with melengestrol acetate. I. Effect of dosage and route of administration. *J. Reprod. Fertil. (Suppl.1):*185.
- Zimbelman, R. G., J. W. Lauderdale, J. H. Sokolowski, and T. G. Schalk. 1970. Safety and pharmacologic evaluations of melengestrol acetate in cattle and other animals. A review. *J.A.V.M.A.* 157:1528-1536.